

ab112157

Rhod-4 No Wash Calcium Assay Kit

Instructions for Use

For detecting calcium in cells without washing by using our proprietary fluorescence probe

This product is for research use only and is not intended for diagnostic use.

Table of Contents

1. Introduction	3
2. Protocol Summary	5
3. Kit Contents	6
4. Storage and Handling	6
5. Additional Materials Required	6
6. Assay Protocol	7
7. Data Analysis	10
8. Troubleshooting	11

1. Introduction

Abcam Rhod-4 No Wash Calcium Assay Kit (ab112157) is a fluorescence-based assay for detecting intracellular calcium mobilization. Cells expressing a GPCR of interest that signals through calcium are pre-loaded with Rhod-4 dye, which can cross cell membranes. Once inside the cell, the lipophilic blocking groups of Rhod-4 are cleaved by non-specific cell esterases, resulting in a negatively charged fluorescent dye which stays inside the cells. Its fluorescence is greatly enhanced upon binding to calcium. When cells are stimulated with screening compounds, the receptor signals release of intracellular calcium, which significantly increase the fluorescence of Rhod-4. The characteristics of its long wavelength, high sensitivity, and >250 times fluorescence increases make Rhod-4 the brightest red calcium indicator available in the market, and it is ideal tool for the measurement of cellular calcium through HTS screening.

ab112156 provides an optimized assay method for monitoring the G-protein-coupled receptors and calcium channels. It can be performed in a convenient 96-well or 384-well microtiter-plate format and easily adapted to automation. Compared to Fluo-8, Rhod-4 is more photostable, making its fluorescence imaging more robust.

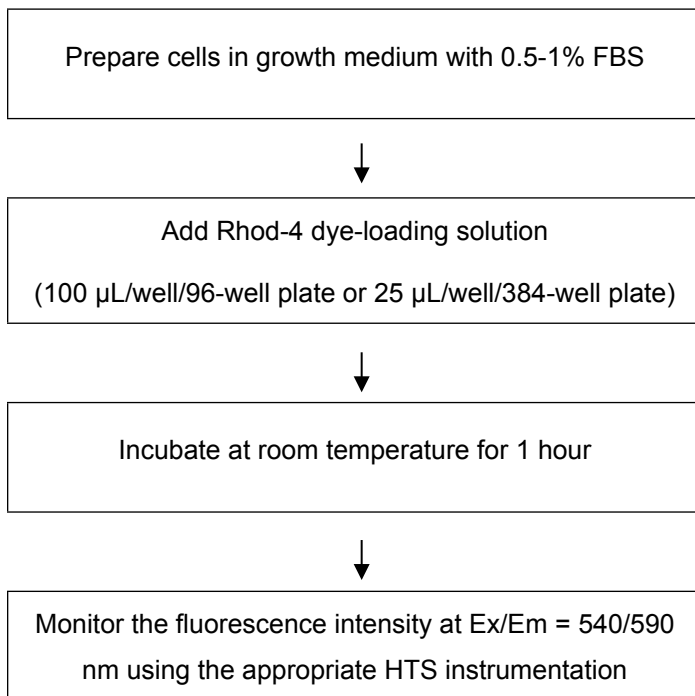
This product has been optimized for HTS screening. We recommend to use this product with the following imaging plate readers: FLIPR™, FDSS, BMG NOVOstar™, FlexStation, ViewLux, IN Cell Analyzer or Arrayscan.

Kit Key Features

- **Longer Wavelengths:** Multiple excitations at 488, 514, 532 and 546 nm; maximum emission at ~555 nm.
- **Larger Assay Window:** 2 fold brighter and 10 fold larger assay window than that of other probes.
- **Convenient:** Formulated to have minimal hands-on time. No wash required.
- **Versatile Applications:** Compatible with many cell lines and receptors.

2. Protocol Summary

Summary for One 96-well Plate; Warning: Do not add additional probenecid.



Note: Thaw all the kit components to room temperature before starting the experiment.

3. Kit Contents

Components	Amount (1 plate)	Amount (10 Plate)
Component A: Rhod-4	1 vial	1 vial
Component B: 10X Pluronic® F127 Plus	1 mL	10 mL
Component C: HHBS	9 mL	100 mL

4. Storage and Handling

Keep at -20°C. Avoid exposure to light.

5. Additional Materials Required

- A 96 or 384-well microplate: A tissue culture microplate with black wall and clear bottom.
- A HTS fluorescence microplate reader with a filter set of Ex/Em = 488 to 545/555 to 590 nm (optimal Ex/Em = 540/590 nm). We recommend the following instruments: FLIPR™, FDSS, BMG NOVOstar™, FlexStation, ViewLux, IN Cell Analyzer or Arrayscan.
- HHBS (1X Hank's with 20 mM Hepes Buffer, pH 7.0).
- 100% DMSO.

- (Optional) Carbachol (ab141354) or other GPCR agonists to induce agonist-mediated calcium release in cells.

6. Assay Protocol

Note: *This protocol is for one 96 - well plate.*

Warning: *Do not add additional probenecid.*

A. Prepare Cells

1. For adherent cells: Plate cells overnight in growth medium at 40,000 to 80,000 cells/well/100 μ L for a 96-well plate or 10,000 to 20,000 cells/well/25 μ L for a 384-well plate.
2. For non-adherent cells: Centrifuge the cells from the culture medium and then suspend the cell pellet in Rhod-4 dye-loading solution (see Step B.4) at 125,000 to 250,000 cells/well/100 μ L for a 96-well poly-D lysine plate or 30,000 to 60,000 cells/well/25 μ L for a 384-well poly-D lysine plate. Centrifuge the plate at 800 rpm for 2 minutes with brake off prior to the experiments.

Note: Each cell line should be evaluated on an individual basis to determine the optimal cell density for the intracellular calcium mobilization.

B. Prepare Rhod-4 Dye-loading Solution

1. Thaw all the kit components at room temperature before use.
2. Make Rhod-4 stock solution: Add 20 μ L of DMSO into the vial of Rhod-4 (Component A), and mix them well.

Note: 20 μ L of Rhod-4 stock solution is enough for one plate. Unused Rhod-4 stock solution can be aliquoted and stored at < -20 °C for more than one month if the tubes are sealed tightly. Protect from light and avoid repeated freeze-thaw cycles.

3. Make 1X assay buffer: Add 9 mL of HHBS (Component C) into the bottle of 10X Pluronic® F127 Plus (1 mL, Component B), and mix them well.

Note: 10 mL of 1X assay buffer is enough for one plate. Aliquot and store un-used 1X assay buffer at < -20 °C. Protect from light and avoid repeated freeze-thaw cycles.

4. Make Rhod-4 dye-loading solution for one cell plate: Add 20 μ L of Rhod-4 stock solution (from Step 2) into 10 mL of 1X assay buffer (from Step 3), and mix them well. This working solution is stable for at least 2 hours at room temperature.

C. Run Calcium Assay:

1. Add 100 μL /well (96-well plate) or 25 μL /well (384-well plate) of Rhod-4 dye-loading solution (from Step B.4) into the cell plate.

Note: Alternatively, grow the cells in growth medium with 5-10% FBS to improve cell growth. In this case, it is important to replace the growth medium with HHBS buffer in order to minimize background fluorescence, and compound interference with serum

2. Incubate the dye-loading plate in a cell incubator for 30 minutes, and then incubate the plate at room temperature for another 30 minutes.

Note 1: If the assay requires 37 °C, perform the experiment immediately without further room temperature incubation.

Note 2: If the cells can function well at room temperature for longer time, incubate the cell plate at room temperature for 1-2 hours.

3. Prepare the compound plate with HHBS or your desired buffer.
4. Using your instrument that contains a pipettor (such as Flexstation from Molecular Devices), dispense compounds directly onto to the cell plate while collecting

the data simultaneously (see next step for Ex/Em instructions).

Note: Typically the instrument collects the data every second for 100 seconds. Use the max signal to generate the plot. You can use a fluorescence microscope by adding the stimuli while taking the picture simultaneously.

5. Run the calcium flux assay by monitoring the fluorescence intensity at Ex/Em = 540/590 nm.

7. Data Analysis

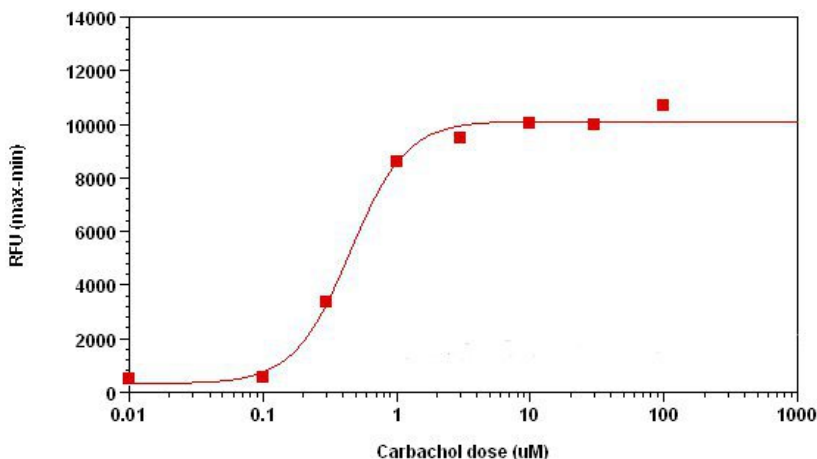


Figure 1. Carbachol Dose Response was measured in HEK-293 cells with ab112157. HEK-293 cells were seeded overnight at 40,000

cells/100 μL /well in a black wall/clear bottom 96-well plate. The cells were incubated with 100 μL of dye-loading solution using ab112157 for 1 hour at room temperature. Carbachol (25 μL /well) was to achieve the final indicated concentrations. The EC_{50} of Rhod-4 is about 0.6 μM .

8. Troubleshooting

Problem	Reason	Solution
Assay not working	Assay buffer at wrong temperature	Assay buffer must not be chilled - needs to be at RT
	Protocol step missed	Re-read and follow the protocol exactly
	Plate read at incorrect wavelength	Ensure you are using appropriate reader and filter settings (refer to datasheet)
	Unsuitable microtiter plate for assay	Fluorescence: Black plates (clear bottoms); Luminescence: White plates; Colorimetry: Clear plates. If critical, datasheet will indicate whether to use flat- or U-shaped wells
Unexpected results	Measured at wrong wavelength	Use appropriate reader and filter settings described in datasheet
	Samples contain impeding substances	Troubleshoot and also consider deproteinizing samples
	Unsuitable sample type	Use recommended samples types as listed on the datasheet
	Sample readings are outside linear range	Concentrate/ dilute samples to be in linear range

Problem	Reason	Solution
Samples with inconsistent readings	Unsuitable sample type	Refer to datasheet for details about incompatible samples
	Samples prepared in the wrong buffer	Use the assay buffer provided (or refer to datasheet for instructions)
	Samples not deproteinized (if indicated on datasheet)	Use the 10kDa spin column (ab93349) or appropriate deproteinization protocol
	Cell/ tissue samples not sufficiently homogenized	Increase sonication time/ number of strokes with the Dounce homogenizer
	Too many freeze-thaw cycles	Aliquot samples to reduce the number of freeze-thaw cycles
	Samples contain impeding substances	Troubleshoot and also consider deproteinizing samples
	Samples are too old or incorrectly stored	Use freshly made samples and store at recommended temperature until use
Lower/ Higher readings in samples and standards	Not fully thawed kit components	Wait for components to thaw completely and gently mix prior use
	Out-of-date kit or incorrectly stored reagents	Always check expiry date and store kit components as recommended on the datasheet
	Reagents sitting for extended periods on ice	Try to prepare a fresh reaction mix prior to each use

	Incorrect incubation time/ temperature	Refer to datasheet for recommended incubation time and/ or temperature
	Incorrect amounts used	Check pipette is calibrated correctly (always use smallest volume pipette that can pipette entire volume)
Standard curve is not linear	Not fully thawed kit components	Wait for components to thaw completely and gently mix prior use
	Pipetting errors when setting up the standard curve	Try not to pipette too small volumes
	Incorrect pipetting when preparing the reaction mix	Always prepare a master mix
	Air bubbles in wells	Air bubbles will interfere with readings; try to avoid producing air bubbles and always remove bubbles prior to reading plates
	Concentration of standard stock incorrect	Recheck datasheet for recommended concentrations of standard stocks
	Errors in standard curve calculations	Refer to datasheet and re-check the calculations
	Use of other reagents than those provided with the kit	Use fresh components from the same kit

For all technical and commercial enquires please go to:

www.abcam.com/contactus

www.abcam.cn/contactus (China)

www.abcam.co.jp/contactus (Japan)

Copyright © 2025 Abcam, All Rights Reserved. The Abcam logo is a registered trademark.

All information / detail is correct at time of going to print.