

# **ab115050 – Histone H3 (methyl K27) Assay Kit**

Instructions for Use

For the measurement of the global methylation of histone H3K27

This product is for research use only and is not intended for diagnostic use.

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## 1. BACKGROUND

Epigenetic activation or inactivation of genes plays a critical role in many important human diseases, especially in cancer. A major mechanism for epigenetic inactivation of the genes is methylation of CpG islands in genome DNA caused by DNA methyltransferases. Histone methyltransferases (HMTs) control or regulate DNA methylation through chromatin-dependent transcription repression or activation. HMTs transfer 1-3 methyl groups from S-adenosyl-L-methionine to the lysine and arginine residues of histone proteins. G9a and polycomb group enzyme such as EZH2 are histone methyltransferases that catalyze Methylation of histone H3 at lysine 27 (H3K27) in mammalian cells. H3K27 methylation mediates heterochromatin formation by forming a binding site for polycomb and also participates in silencing gene expression at euchromatic sites. Increased global H3K27 methylation is also found to be involved in some pathological processes such as cancer progress. There are only a couple of methods such as western blot used for measuring histone H3K27 methylation. The methods available so far are time consuming and labor intensive, or have low throughput. The Histone H3 (methyl K27) Assay Kit addresses these problems by using a unique procedure to measure global methylation of histone H3K27.

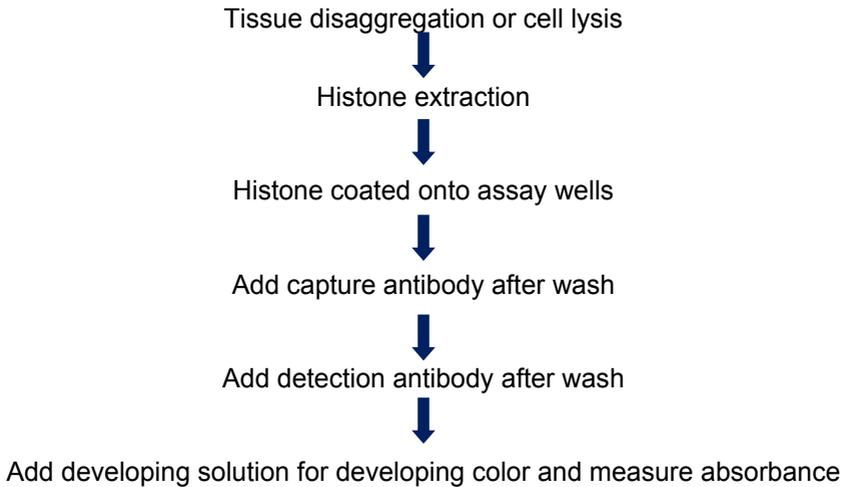
The kit has the following features:

- Quick and efficient procedure, which can be finished within 5 hours
- Innovative colorimetric assay without the need for radioactivity, electrophoresis, or chromatography
- Strip microplate format makes the assay flexible: manual or high throughput
- Simple, reliable, and consistent assay conditions

The Histone H3 (methyl K27) Assay Kit is designed for measuring global histone H3K27 methylation. In an assay with this kit, the histone proteins are stably spotted on the strip wells. The methylated histone H3K27 can be recognized with a high-affinity antibody. The ratio or amount of methylated

H3K27 can be quantified through HRP conjugated secondary antibody-color development system and is proportional to the intensity of color development.

## 2. ASSAY SUMMARY



### **3. PRECAUTIONS**

**Please read these instructions carefully prior to beginning the assay.**

All kit components have been formulated and quality control tested to function successfully as a kit. Modifications to the kit components or procedures may result in loss of performance.

### **4. STORAGE AND STABILITY**

**Store kit as given in the table upon receipt away from light.**

Observe the storage conditions for individual prepared components in sections 9 & 10.

For maximum recovery of the products, centrifuge the original vial prior to opening the cap.

Check if the 10X Wash Buffer contains salt precipitates before use. If so, warm at room temperature or 37°C and shake the buffer until the salts are re-dissolved

## 5. MATERIALS SUPPLIED

Item	48 Tests	96 Tests	Storage Condition (Before Preparation)
10X Lysis Buffer	5 mL	10 mL	RT
Extraction Buffer	8 mL	16 mL	RT
10X Wash Buffer	14 mL	28 mL	4°C
Histone Buffer	0.5 mL	1 mL	RT
Blocking Buffer	10 mL	20 mL	4°C
Antibody Buffer	6 mL	12 mL	RT
Capture Antibody, 100 µg/mL	25 µL	50 µL	4°C
Detection Antibody, 400 µg/mL	10 µL	20 µL	-20°C
Developing Solution	5 mL	10 mL	4°C
Stop Solution	3 mL	6 mL	RT
Methylated H3K27 Control, 60 µg/mL*	10 µL	20 µL	-20°C
8-Well Assay Strips (with frame)	6	12	4°C

\*Standard Control Strips are identified by a green ring around the top.

## 6. MATERIALS REQUIRED, NOT SUPPLIED

These materials are not included in the kit, but will be required to successfully utilize this assay:

- Orbital shaker
- Pipettes and pipette tips
- Microplate reader
- 1.5 mL microcentrifuge tubes
- 60 or 100 mm plate
- Dounce homogenizer
- 100% TCA solution
- Glycerol
- Acetone
- 5% HCl
- Distilled water

## 7. LIMITATIONS

- Assay kit intended for research use only. Not for use in diagnostic procedures
- Do not use kit or components if it has exceeded the expiration date on the kit labels
- Do not mix or substitute reagents or materials from other kit lots or vendors. Kits are QC tested as a set of components and performance cannot be guaranteed if utilized separately or substituted
- Any variation in operator, pipetting technique, washing technique, incubation time or temperature, and kit age can cause variation in binding

## 8. TECHNICAL HINTS

- Avoid foaming or bubbles when mixing or reconstituting components
- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions
- Ensure plates are properly sealed or covered during incubation steps
- Complete removal of all solutions and buffers during wash steps
- **This kit is sold based on number of tests. A ‘test’ simply refers to a single assay well. The number of wells that contain sample, control or standard will vary by product. Review the protocol completely to confirm this kit meets your requirements. Please contact our Technical Support staff with any questions**

## 9. REAGENT PREPARATION

Prepare fresh reagents immediately prior to use.

### 9.1. **1X Lysis Buffer**

Dilute 10X Wash Buffer with distilled water (pH 7.2-7.5) at a 1:10 ratio (e.g. 1 mL of Lysis Buffer + 9 mL of distilled water).

### 9.2. **1X Wash Buffer**

Dilute 10X Wash Buffer with distilled water (pH 7.2-7.5) at a 1:10 ratio (e.g. 1 mL of Wash Buffer + 9 mL of distilled water).

### 9.3. **Diluted Capture Antibody**

Dilute Capture Antibody (at a 1:100 ratio) to 1 µg/mL with Antibody buffer.

### 9.4. **Diluted Detection Antibody**

Dilute Detection Antibody (at a 1:1000 ratio) to 0.4 µg/mL with Antibody Buffer.

## 10. SAMPLE PREPARATION

### 10.1. **Preparation of Nucleic Extracts:**

10.1.1. **For tissue samples:** Place the tissue sample into a 60 or 100 mm plate. Remove unwanted tissue such as fat and necrotic material from the sample. Weigh the sample and cut it into small pieces (1-2 mm<sup>3</sup>) with a scalpel or scissors.

10.1.2. Transfer tissue pieces to a Dounce homogenizer. Add 1 mL of the 1X Lysis Buffer per every 200 mg of tissue and disaggregate tissue pieces by 10-30 strokes.

10.1.3. Transfer homogenized mixture to a 15 mL conical tube and centrifuge at 3000 rpm for 5 minutes at 4°C. If total mixture volume is less than 2 mL, transfer mixture to a 2 mL vial and centrifuge at 10,000 rpm for 1 minute at 4°C. Remove supernatant.

10.1.4. **For Adherent Cells:** Cells (treated or untreated) are grown to 70-80% confluency, then trypsinized and collected into a 15 mL conical tube. Count cells in a hemocytometer.

- 10.1.5. Centrifuge the cells at 1000 rpm for 5 minutes and discard the supernatant. Wash cells with 10 mL of PBS once by centrifugation at 1000 rpm for 5 minutes. Discard the supernatant.
  - 10.1.6. Add the 1X Lysis Buffer to re-suspend cell pellet (200  $\mu\text{L}/1 \times 10^6$  cells). Transfer cell suspension to a 1.5 mL vial and incubate on ice for 5 minutes and vortex occasionally.
  - 10.1.7. Pellet cell debris by centrifuging at 12,000 rpm for 30 seconds.
  - 10.1.8. **For Suspension Cells:** Collect cells (treated or untreated) into a 15 mL conical tube (1-2 $\times 10^6$  cells are required for each reaction). Count cells in a hemocytometer.
  - 10.1.9. Centrifuge the cells at 1000 rpm for 5 minutes and discard the supernatant. Wash cells with 10 mL of PBS once by centrifugation at 1000 rpm for 5 minutes. Discard the supernatant.
  - 10.1.10. Add the 1X Lysis Buffer to re-suspend cell pellet (100  $\mu\text{L}/1 \times 10^6$  cells). Transfer cell suspension to a 1.5 mL vial and incubate on ice for 5 minutes and vortex occasionally. Pellet cell debris by centrifuging at 12,000 rpm for 30 seconds.
- 10.2. **Preparation of Histone Extract:**
- 10.2.1. Add glycerol to Extraction Buffer at a 1:10 ratio (e.g. add 100  $\mu\text{L}$  of glycerol to 900  $\mu\text{L}$  of Extraction Buffer) to prepare the Extraction Buffer/Glycerol solution. Add the 1X Lysis Buffer to cell debris (10  $\mu\text{L}/1 \times 10^6$  cells or 40 mg of tissue), followed by adding 3 volumes of Extraction Buffer/Glycerol solution. Mix by vortex and incubate on ice for 5 minutes.
  - 10.2.2. Pellet nucleic debris by centrifuging at 12,000 rpm for 5 minutes at 4°C. Transfer the supernatant to a 1.5 mL vial.
  - 10.2.3. Add 100% TCA solution to the supernatant at a 1:4 ratio (e.g. add 100  $\mu\text{L}$  of TCA to 300  $\mu\text{L}$  of supernatant; final concentration of TCA should be 25%). Incubate on ice for 30 minutes.

- 10.2.4. Collect the precipitate by centrifuging at 12,000 rpm for 10 minutes at 4°C.
- 10.2.5. Remove supernatant and add 1 mL of acetone containing 0.1% HCl to precipitate. Mix and incubate on ice for 1 minute.
- 10.2.6. Collect the pellet by centrifuging at 12,000 rpm for 2 minutes at 4°C. Wash the pellet with 1 mL of acetone. Allow 1 minute on ice for wash.
- 10.2.7. Collect the pellet by centrifuging at 12,000 rpm for 2 minutes at 4°C. Remove supernatant as much as possible and air dry the pellet for 5 minutes.
- 10.2.8. Add distilled water to dissolve pellet (10  $\mu$ L of water per amount of pellet extracted from  $1 \times 10^6$  cells or 40 mg of tissue) and measure histone protein concentration.

Histone extracts can be used immediately or stored at -20°C for future use.

## 11. ASSAY PROCEDURE

- 11.1 Determine the number of strip wells required. Leave these strips in the plate frame (remaining unused strips can be placed back in the bag. Seal the bag tightly and store at 4°C).
- 11.2 Adjust protein concentration to 200 ng/μL or 400 ng/μL with Histone Buffer and add 5 μL (1-2 μg) of the protein solution into the central area of each well. Spread out the solution over the bottom of the strip well by pipetting the solution up and down several times, and incubate at 37°C (with no humidity) for 60-90 minutes to evaporate the solution and dry the wells. For the blank, add 5 μL of Histone Buffer to the wells. For the positive control, dilute Methylated H3K27 Control to 2-30 ng/μL with Histone Buffer, and then add 5 μL (10-150 ng) of the diluted Methylated H3K27 Control solution to the wells.
- 11.3 Add 150 μL of Blocking Buffer to the dried wells and incubate at 37°C for 30-45 minutes.
- 11.4 Aspirate and wash the wells with 150 μL of 1X Wash Buffer three times.
- 11.5 Add 50 μL of the Diluted Capture Antibody to the wells and incubate at room temperature for 60 minutes on an orbital shaker (50-100 rpm).
- 11.6 Aspirate and wash the wells with 150 μL of 1X Wash Buffer four times.
- 11.7 Add 50 μL of the Diluted Detection Antibody to the wells and incubate at room temperature for 30 minutes.
- 11.8 Aspirate and wash the wells with 150 μL of the 1X Wash Buffer five times.
- 11.9 Add 100 μL of Developing Solution to the wells and incubate at room temperature for 2-10 minutes away from light. Monitor the color development in the sample and standard wells (blue).
- 11.10 Add 50 μL of Stop Solution to the wells and read absorbance on a microplate reader at 450 nm.

## 12. ANALYSIS

Calculate % H3K27 methylation:

$$\text{Methylation \%} = \frac{\text{Treated (Tested) Sample OD} - \text{Blank OD}}{\text{Untreated (Control) Sample OD} - \text{Blank OD}} \times 100\%$$

For amount quantification plot delta OD values (positive control OD – negative control OD) versus amount of Standard Control added in the wells and determine the slope as delta OD/ng.

Calculate the amount of methylated H3K27 using the following formula:

$$\text{Amount (ng/mg protein)} = \frac{\text{Sample OD} - \text{Blank OD}}{\text{Protein } (\mu\text{g})^* \times \text{Slope}} \times 1000$$

\*Histone extract amount added into the sample well at step 11.2.

13. TROUBLESHOOTING

<b>Problem</b>	<b>Cause</b>	<b>Solution</b>
No Signal for Both the Standard Control and the Samples	Reagents are added incorrectly	Check if reagents are added in order and if some steps of the procedure are omitted by mistake
	Incubation time and temperature is incorrect	Ensure the incubation time and temperature described in the protocol is followed correctly
No Signal or Very Weak Signal for Only the Standard Control	The amount of Standard Control is not added into the "standard control wells or is added insufficiently	Ensure a sufficient amount of control is added to the well
	The positive control is degraded due to incorrect storage.	Follow the guidance in the protocol for storage of positive control
No Signal for Only the Sample	The protein sample is not properly extracted	Follow the protocol instructions for the histone protein extraction
	The protein amount is added into well insufficiently	Ensure extract contains a sufficient amount of protein
	Protein extracts are incorrectly stored	Ensure the protein extracts are stored at -20°C or -80°C
High Background Present for the Blank	The well is not washed enough	Check if wash at each step is performed according to the protocol
	Contaminated by the Standard control	Ensure the well is not contaminated from adding the control protein or by using control protein contaminated tips
	Overdevelopment	Decrease development time in step 11.9

14. NOTES

# RESOURCES

**UK, EU and ROW**

Email: [technical@abcam.com](mailto:technical@abcam.com) | Tel: +44-(0)1223-696000

**Austria**

Email: [wissenschaftlicherdienst@abcam.com](mailto:wissenschaftlicherdienst@abcam.com) | Tel: 019-288-259

**France**

Email: [supportscientifique@abcam.com](mailto:supportscientifique@abcam.com) | Tel: 01-46-94-62-96

**Germany**

Email: [wissenschaftlicherdienst@abcam.com](mailto:wissenschaftlicherdienst@abcam.com) | Tel: 030-896-779-154

**Spain**

Email: [soportecientifico@abcam.com](mailto:soportecientifico@abcam.com) | Tel: 911-146-554

**Switzerland**

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**US and Latin America**

Email: [us.technical@abcam.com](mailto:us.technical@abcam.com) | Tel: 888-77-ABCAM (22226)

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