

ab126437 - EGFR (pY1045) + total EGFR Human ELISA Kit

Instructions for Use

For the quantitative measurement of human, phosphorylated EGFR (Tyr 1045) and total EGFR concentrations in cell lysates.

This product is for research use only and is not intended for diagnostic use.

Table of Contents

1. Introduction	3
2. Assay Summary	4
3. Kit Contents	5
4. Storage and Handling	6
5. Additional Materials Required	7
6. Preparation of Samples	8
7. Preparation of Reagents	9
8. Assay Method	12
9. Data Analysis	14
10. Troubleshooting	16

1. Introduction

ab126437 is a very rapid, convenient and sensitive assay kit that can monitor the activation or function of important biological pathways in cell lysates. By determining phosphorylated EGFR protein in your experimental model system, you can verify pathway activation in your cell lysates. You can simultaneously measure numerous different cell lysates without spending excess time and effort in performing a Western Blotting analysis.

This sandwich ELISA kit is an in vitro enzyme-linked immunosorbent assay for the measurement of human phospho-EGFR (Tyr1045) and total EGFR (help normalize the results of phospho-EGFR from different cell lysate being compared). An anti-EGFR antibody has been coated onto a 96-well plate. Samples are pipetted into the wells and phosphorylated and total EGFR present in a sample is bound to the wells by the immobilized antibody. The wells are washed and anti-EGFR (Tyr1045) or anti-total-EGFR antibody is used to detect phosphorylated or non-phosphorylated EGFR. After washing away unbound antibody, HRP-conjugated anti-rabbit IgG or HRP-Streptavidin is pipetted to the wells. The wells are again washed, a TMB substrate solution is added to the wells and color develops in proportion to the amount of EGFR (Tyr1045) or total EGFR bound. The Stop Solution changes the color from blue to yellow, and the intensity of the color is measured at 450 nm.

2. Assay Summary

Prepare all reagents, samples and standards as instructed.



Add 100 μ l sample or positive control to each well. Incubate 2.5 hours at room temperature or overnight at 4°C.



Add 100 μ l prepared primary antibody to each well. Incubate 1.5 hours at room temperature.



Add 100 μ l prepared secondary antibody solution. Incubate for overnight (corresponding well of adding Rabbit phospho-EGFR) or incubate for 1 hour at room temperature (corresponding well of adding HRP-Streptavidin)



Add 100 μ l TMB One-Step Substrate Reagent to each well. Incubate 30 minutes at room temperature.



Add 50 μ l Stop Solution to each well. Read at 450 nm immediately.

3. Kit Contents

- EGFR Microplate (Item A): 96 wells (12 strips x 8 wells) coated with monoclonal anti-EGFR antibody.
- Wash Buffer Concentrate (20x) (Item B): 25 mL
- Anti-phospho-EGFR (Tyr 1045) (Item C): 1 vial
- HRP-conjugated Anti-rabbit IgG (Item D), 25 μ L
- Biotinylated Anti-EGFR - Goat anti-human EGFR (Item L): 1 vial
- 600X HRP-Streptavidin concentrate (Item G): 200 μ L
- 5X Assay Diluent (Item E): 15 mL. For diluting cell lysate sample, Biotinylated antibody (Item L) and HRP- Streptavidin (Item G) diluent.
- TMB One-Step Substrate Reagent (Item H): 12 mL
- Stop Solution (Item I): 8 mL
- 2X Cell Lysis Buffer (Item J): 5 mL
- Positive Control - Lyophilized powder from A431 cell lysate (Item K): 1 vial

4. Storage and Handling

Upon receipt, the kit should be stored at -20°C . After initial use, Wash Buffer Concentrate (Item B), HRP-conjugated Anti-rabbit IgG (Item D), Assay Diluent (Item E), TMB One-Step Substrate Reagent (Item H), Stop Solution (Item I) and Cell Lysis Buffer (Item J) and Biotinylated Antibody should be stored at 4°C to avoid repeated freeze-thaw cycles. Anti-phospho-EGFR (Tyr 1045) (Item C) and HRP-Streptavidin should be stored at -20°C . Return unused wells to the pouch containing desiccant pack, reseal along entire edge and store at -20°C . Reconstituted Positive Control (Item K) should be stored at -70°C .

5. Additional Materials Required

- Microplate reader capable of measuring absorbance at 450 nm.
- Protease and Phosphatase inhibitors.
- Shaker.
- Precision pipettes to deliver 2 μ l to 1 ml volumes.
- Adjustable 1-25 ml pipettes for reagent preparation.
- 100 ml and 1 L graduated cylinders.
- Distilled or deionized water.
- Tubes to prepare sample dilutions.

6. Preparation of Samples

Cell lysates - Rinse cells with PBS, making sure to remove any remaining PBS before adding the Cell Lysis Buffer. Solubilize cells at 4×10^7 cells/ml in 1x Cell Lysis Buffer (we recommend adding protease and phosphatase inhibitors to Cell Lysis Buffer prior to sample preparation). Pipette up and down to resuspend and incubate the lysates with shaking at 2 - 8°C for 30 minutes. Microcentrifuge at 13,000 rpm for 10 minutes at 2 - 8°C, and transfer the supernates into a clean test tube. Lysates should be used immediately or aliquoted and stored at -70°C. Avoid repeated freeze-thaw cycles. Thawed lysates should be kept on ice prior to use.

For the initial experiment, we recommend to do a serial dilution testing such as 5-fold and 50-fold dilution for your cell lysates with 1x Assay Diluent (Item E) before use.

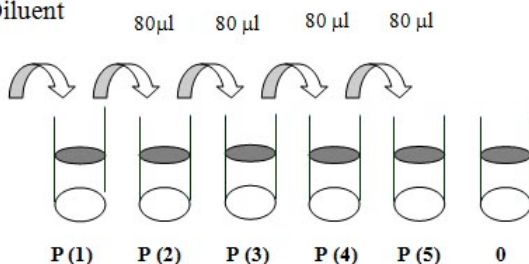
Note: The fold dilution of sample used depends on the abundance of phosphorylated proteins and should be determined empirically. More of the sample can be used if signals are too weak. If signals are too strong, the sample can be diluted further.

Cell Lysis Buffer should be diluted 2-fold with deionized or distilled water before use (recommend to add protease and phosphatase inhibitors).

7. Preparation of Reagents

1. Bring all reagents and samples to room temperature (18 - 25°C) before use.
2. Item E, Assay Diluent should be diluted 5-fold with deionized or distilled water before use.
3. Preparation of Positive Control: Briefly spin the Positive Control vial of Item K. Add 400 μ l 1x Assay Diluent (Item E, Assay Diluent should be diluted 5-fold with deionized or distilled water before use) into Item K vial to prepare Positive Control Solution (P-1) (See i. Positive Control of Data Analysis for a typical result). Dissolve the powder thoroughly by a gentle mix. Pipette 320 μ l 1x Assay Diluent into each tube. Add 80 μ l prepared P-1 Solution from the vial of Item K, into a tube with 320 μ l 1x Assay Diluent to produce a dilution series (shown below). Mix each tube thoroughly before the next transfer. 1x Assay Diluent serves as the background.

400 μ l 1x Assay Diluent



4. If the Wash Concentrate (20x) (Item B) contains visible crystals, warm to room temperature and mix gently until dissolved. Dilute 20 ml of Wash Buffer Concentrate into deionized or distilled water to yield 400 ml of 1x Wash Buffer.
5. Briefly spin the anti-phospho-EGFR (Tyr 1045) (Item C) before use. Pipette up and down to mix gently. The anti-phospho-EGFR (Tyr 1045) should be diluted 1,000-fold with 1x Assay Diluent. For example, add 5 μ l anti-phospho-EGFR (Tyr 1045) into a tube with 5.0 ml 1x Assay Diluent to prepare a 1,000-fold diluted antibody.
6. Briefly spin the HRP-conjugated anti-rabbit IgG (Item D) before use. Pipette up and down to mix gently. HRP-conjugated anti-rabbit IgG concentrate should be diluted 500-fold with 1x Assay Diluent.
7. Briefly spin the Detection Antibody vial (Item L) before use. Add 100 μ l of 1x Assay Diluent into the vial to prepare a detection antibody concentrate. Pipette up and down to mix gently (the

concentrate can be stored at 4°C for 5 days. It can be used within one month if stored at -80°C. Avoid repeated freeze-thaw cycles). The detection antibody concentrate should be diluted 200-fold with 1x Assay Diluent and used in step 4 of Assay Method.

8. Briefly spin the HRP-Streptavidin concentrate vial (Item G) and pipette up and down to mix gently before use since precipitation may form during storage. HRP-Streptavidin concentrate should be diluted 600-fold with 1x Assay Diluent.

For example: Briefly spin the vial (Item G) and pipette up and down to mix gently. Add 20 μ l of HRP-Streptavidin concentrate into a tube with 12 ml 1x Assay Diluent B to prepare a 600-fold diluted HRP-Streptavidin solution (don't store the diluted solution for next day use). Mix well.

9. Cell Lysis Buffer should be diluted 2-fold with deionized or distilled water before use (recommend to add protease and phosphatase inhibitors).

8. Assay Method

1. Bring all reagents to room temperature (18 - 25°C) before use. It is recommended that all samples or Positive Control should be run at least in duplicate.
2. Add 100 µl of each sample or positive control into appropriate wells. Cover well with plate holder and incubate for 2.5 hours at room temperature or overnight at 4°C with shaking.
3. Discard the solution and wash 4 times with 1x Wash Solution. Wash by filling each well with Wash Buffer (300 µl) using a multi-channel pipette or autowasher. Complete removal of liquid at each step is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating or decanting. Invert the plate and blot it against clean paper towels.
4. Add 100 µl of 1,000-fold diluted anti-phospho-EGFR (Tyr1045) (Preparation of Reagents step 5) to corresponding well for detecting phospho-EGFR or 100 µl of diluted Biotinylated anti-EGFR (see Preparation of Reagents step 7) to corresponding well (help normalize the results of phospho-EGFR from different cell lysate being compared) for detecting total EGFR. Incubate for 1.5 hour at room temperature with shaking.
5. Discard the solution. Repeat the wash as in step 3.

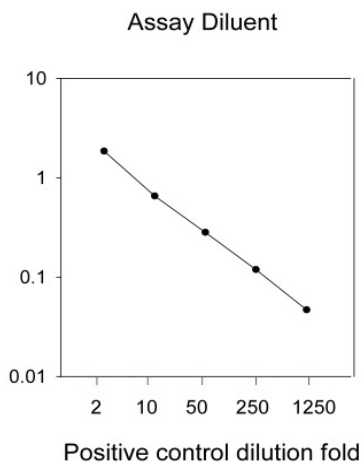
6. Add 100 μ l of 500-fold diluted HRP-conjugated anti-rabbit IgG (see Preparation of Reagents step 6) to detect Rabbit phospho-EGFR (Tyr 1045) (corresponding well of adding Rabbit phospho-EGFR). Incubate for overnight at 4°C. Add 100 μ l of 600 diluted HRP-Streptavidin (see Preparation of Reagents step 8) to detect Biotinylated EGFR antibody (corresponding well of adding Biotinylated anti-EGFR antibody). Incubate for 1 hour at room temperature with shaking.
7. Discard the solution. Repeat the wash as in step 3.
8. Add 100 μ l of TMB One-Step Substrate Reagent (Item H) to each well. Incubate for 30 minutes at room temperature in the dark with shaking.
9. Add 50 μ l of Stop Solution (Item I) to each well. Read at 450nm immediately.

9. Data Analysis

ELISA data analysis: Average the duplicate readings for each sample or positive control then subtract the average blank optical density.

i. Positive Control

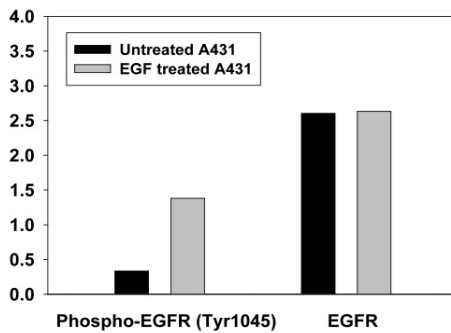
A431 cells were treated with recombinant human EGF at 37°C for 20 min. Solubilize cells at 2×10^7 cells/ml in Cell Lysis Buffer. Serial dilutions of lysates were analyzed in this ELISA. Please see step 3 of Preparation of Reagents for detail.



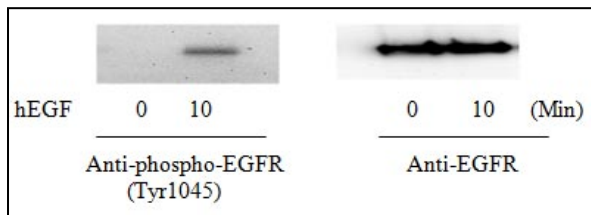
ii. **Recombinant Human EGF Stimulation of A431 Cell Lines**

A431 cells were treated or untreated with 100 ng/ml recombinant human EGF for 10 min. Cell lysates were analyzed using this phosphoELISA and Western Blot.

a) **ELISA**



b) **Western Blot Analysis**



10. Troubleshooting

Problem	Cause	Solution
Sample signals	Too low: Sample concentration is too low.	Increasing sample concentration.
	Too high: Sample concentration is too high.	Reduce sample concentration.
Large CV	Inaccurate pipetting	Check pipettes
High background	Plate is insufficiently washed.	Review the manual for proper washing. If using an automated plate washer, check that all ports are unobstructed.
	Contaminated wash buffer.	Make fresh wash buffer.

Positive Control: Low signal	Improper storage of the ELISA kit.	Upon receipt, the kit should be stored at -20°C. Store the positive control at -70°C after reconstitution.
	Stop solution	Stop solution should be added to each well before measurement and read OD immediately.
	Improper primary or secondary antibody dilution.	Ensure correct dilution.

Technical Support

Copyright © 2024 Abcam. All Rights Reserved. The Abcam logo is a registered trademark. All information / detail is correct at time of going to print.

For all technical or commercial enquiries please go to:

www.abcam.com/contactus

www.abcam.cn/contactus (China)

www.abcam.co.jp/contactus (Japan)