

ab150686 Trichrome Stain (Connective Tissue Stain)

For the histological visualization of collagenous connective tissue fibers in tissue sections.

This product is for research use only and is not intended for diagnostic use

Overview

The Trichrome Stain (Connective Tissue Stain) (ab150686) is intended for use in the histological visualization of collagenous connective tissue fibers in tissue sections. The trichrome stain kit may be used on formalin-fixed, paraffin-embedded. For frozen sections, it will require optimization by the user.

Precautions

Please observe safe laboratory practice and consult the safety datasheet.

For general guidelines, precautions, limitations on the use of our assay kits and general assay troubleshooting tips, particularly for first time users, please consult our guide:

www.abcam.com/assaykitguidelines

For typical data produced using the assay, please see the assay kit datasheet on our website.

Storage and Stability:

Store kit at room temperature immediately on receipt and check below for storage for individual components.

Keep away from open flame and refer to the safety datasheet.

Materials Supplied:

Item	Quantity	Storage temperature (before prep)
Bouin's Fluid	125 mL	RT
Weigert's Iron, Hematoxylin (A)	125 mL	RT
Weigert's Iron, Hematoxylin (B)	125 mL	RT
Biebrich Scarlet/Acid Fuchsin Solution	125 mL	RT
Phosphomolybdic/Phosphotungstic Acid	125 mL	RT
Aniline Blue Solution	125 mL	RT
Acetic Acid Solution (1%)	125 mL	RT

Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully perform this assay:

- Absolute alcohol
- Xylene or xylene substitute

Staining protocol

- Equilibrate all materials and prepared reagents to room temperature just prior to use and gently agitate.

1. Deparaffinize sections if necessary and hydrate in distilled water.
2. Preheat Bouin's Fluid in a water bath to 56-64°C in a fume hood or very well ventilated area.
3. Place slide in preheated Bouin's Fluid for 60 minutes followed by a 10 minute cooling period.
4. Rinse slide in tap water until section is completely clear.
5. Rinse once in distilled water.
6. Mix equal parts of Weigert's (A) and Weigert's (B) and stain slide with working Weigert's Iron Hematoxylin for 5 minutes.
7. Rinse slide in running tap water for 2 minutes.
8. Apply Biebrich Scarlet / Acid Fuchsin Solution to slide for 15 minutes.
9. Rinse slide in distilled water.
10. Differentiate in Phosphomolybdic/Phosphotungstic Acid Solution for 10-15 minutes or until collagen is not red.
11. Without rinsing, apply Aniline Blue Solution to slide for 5-10 minutes.
12. Rinse slide in distilled water.
13. Apply Acetic Acid Solution (1%) to slide for 3-5 minutes.
14. Dehydrate very quickly in 2 changes of 95% Alcohol, followed by 2 changes of Absolute Alcohol.
15. Clear in Xylene or Xylene Substitute and mount in synthetic resin.

Technical Support

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