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ab186028 Cellular Reactive Oxygen Species Detection Assay Kit (Orange Fluorescence)

For the detection of Reactive Oxygen Species (ROS) in live cells.

This product is for research use only and is not intended for diagnostic use.

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1. Overview

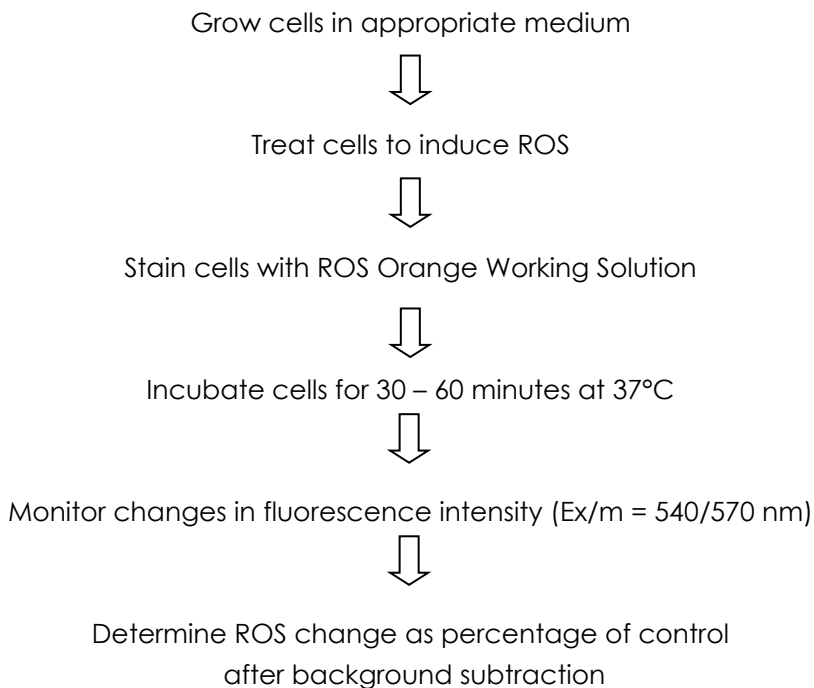
Cellular Reactive Oxygen Species Detection Assay Kit (Orange Fluorescence) (ab186028) is a sensitive fluorometric one-step assay to detect intracellular reactive oxygen species (ROS) such as superoxide and hydroxyl radical in live cells. The assay uses our proprietary ROS Orange dye to quantify ROS: the dye is cell-permeable and reacts with ROS present in the cell to generate an orange fluorescent signal (Ex/Em = 540/570 nm).

The assay can be performed in 1 hour and can be detected by fluorescence microscopy, flow cytometry, microplate fluorometry or high-content imaging. It can be easily adapted to use in 384-well microplate format.

Reactive oxygen species (ROS) are natural by-products of the normal metabolism of oxygen. The two major sources of cellular ROS are Complex I (NADH dehydrogenase ubiquinone-ubiquinol reductase) and Complex III (ubiquinol cytochrome c reductase), both part of the mitochondrial electron transport chain. These two complexes generate ROS particularly when electron transport is slowed by high mitochondrial membrane potential ($\Delta\psi_m$). The major product of ROS in mitochondria is in the form of superoxide and hydroperoxyl radical. Superoxide generated in complex III occurs in the presence of slow electron transport which allows for the ubisemiquinone anion radical to react with oxygen dissolved in the membrane. The exact source of superoxide generated by complex I is less known and it is believed to be due to electron leakage from its iron-sulfur clusters.

Low levels (or optimum levels) of ROS play an important role in signaling pathways. However, when ROS production increases and overwhelms the cellular antioxidant capacity, it can induce macromolecular damage (by reacting with DNA, proteins and lipids) and disrupt thiol redox circuits. In the first instance, damage can lead to apoptosis or necrosis. Disruption of thiol redox circuits can lead to aberrant cell signaling and dysfunctional redox control.

2. Protocol Summary



3. Precautions

Please read these instructions carefully prior to beginning the assay.

- All kit components have been formulated and quality control tested to function successfully as a kit.
- We understand that, occasionally, experimental protocols might need to be modified to meet unique experimental circumstances. However, we cannot guarantee the performance of the product outside the conditions detailed in this protocol booklet.
- Observe good laboratory practices. Gloves, lab coat, and protective eyewear should always be worn. Never pipet by mouth. Do not eat, drink or smoke in the laboratory areas.
- If applicable, please refer to the current Safety Data Sheet (SDS) provided with this product for safety, handling, and disposal information. The most up to date and current versions are available on our website www.abcam.com.

4. Storage and Stability

Store kit at -20°C in the dark immediately upon receipt. Kit has a storage time of 1 year from receipt, providing components have not been reconstituted.

Refer to list of materials supplied for storage conditions of individual components. Observe the storage conditions for individual prepared components in the Materials Supplied section.

Aliquot components in working volumes before storing at the recommended temperature.

5. Limitations

- Assay kit intended for research use only. Not for use in diagnostic procedures.
- Do not mix or substitute reagents or materials from other kit lots or vendors. Kits are QC tested as a set of components and performance cannot be guaranteed if utilized separately or substituted.

6. Materials Supplied

Item	Quantity	Storage Condition (Before prep)	Storage Condition (After prep)
Assay Buffer	20 mL	-20°C	-20°C
DMSO	100 µL	-20°C	-20°C
ROS Orange Dye (lyophilized)	1 vial	-20°C	-20°C

7. Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully perform this assay:

- Fluorometric microplate reader (bottom read mode) or flow cytometer capable of measuring fluorescence at Ex/Em = 540/570 nm
- PBS or HHBS buffer
- Pipettes and pipette tips, including multi-channel pipette
- Assorted glassware for the preparation of reagents and buffer solutions
- Tubes for the preparation of reagents and buffer solutions
- General tissue culture supplies
- Sterile 96 well plate with clear flat bottom, preferably black (if performing assay in microplate format). Use a poly-D-lysine coated plate for suspension cells

8. Technical Hints

- **This kit is sold based on number of tests. A “test” simply refers to a single assay well. The number of wells that contain sample, control or standard will vary by product. Review the protocol completely to confirm this kit meets your requirements. Please contact our Technical Support staff with any questions.**
- Selected components in this kit are supplied in surplus amount to account for additional dilutions, evaporation, or instrumentation settings where higher volumes are required. They should be disposed of in accordance with established safety procedures.
- Avoid foaming or bubbles when mixing or reconstituting components.
- Avoid cross contamination of samples or reagents by changing tips between sample and reagent additions.
- Ensure plates are properly sealed or covered during incubation steps.
- Ensure all reagents and solutions are at the appropriate temperature before starting the assay.
- Make sure all necessary equipment is switched on and set at the appropriate temperature.

9. Reagent Preparation

Briefly centrifuge small vials at low speed prior to opening.

9.1 Assay Buffer:

Ready to use as supplied. Equilibrate to room temperature before use. Store at -20°C.

9.2 DMSO:

Ready to use as supplied. Warm by placing in a 37°C bath for 1 – 5 min to thaw the DMSO solution before use.

9.3 ROS Orange Dye:

Reconstitute dye by adding 40 µL of DMSO (step 9.2) to the vial to prepare **ROS Orange Stain Stock Solution**. Mix well by pipetting up and down. Equilibrate Stock Solution to room temperature before use.

Aliquot unused Stock Solution so that you have enough volume to perform the desired number of assay. Store at -20°C with the cap sealed tightly away from light. Avoid freeze-thaw cycles. Use within 2 months.

10. Assay Procedure – Microplate format

- Equilibrate all materials and prepared reagents to room temperature prior to use.
- We recommend that you assay all controls and samples in duplicate.
- Prepare all reagents and samples as directed in the previous sections.
- Each cell line should be evaluated on an individual basis to determine the optimal cell density.
- The protocol described here is for 96-well microplate format. You can adapt the protocol for 384-well format by dividing working volumes by half.

10.1 Grow cells:

10.1.1 Adherent cells: plate cells overnight in growth medium at $10^4 - 4 \times 10^4$ cells/90 μ L per well.

Δ Note: for 384 well plate, use $2.5 \times 10^3 - 10^4$ cells/20 μ L per well.

10.1.2 Suspension cells: on the day of the assay, centrifuge cells from the culture medium and resuspend the cell pellet in culture medium at $5 \times 10^4 - 10^5$ cells/90 μ L per cell in a poly-D-lysine coated plate.

Δ Note: for 384 well plate, use $10^4 - 2.5 \times 10^4$ cells/20 μ L per well.

10.1.3 Centrifuge plate at 800 rpm for 2 minutes with brake off.

10.2 Prepare ROS Orange Stain working solution:

10.2.1 Add 20 μ L of ROS Orange Stock Solution (Step 9.3) to 10 mL of Assay Buffer and mix well.

Δ Note: ROS Orange Stain working solution is stable for at least 2 hours at room temperature.

10.3 Run ROS assay:

10.3.1 Treat cells with 10 μ L of 10X test compounds in PBS or HHBS. For untreated cells, add 10 μ L of compound buffer.

Δ Note: for 384 well plate, add 5 μ L of 5X test compounds.

10.3.2 Induce ROS production by incubating cell plate at room temperature or in a 37°C/5% CO₂ incubator for at least 15 minutes, or required time for your compound.

Δ Note: for example, incubate HeLa cells for 30 minutes when treated with 100 μ M TBHP (tert-butyl-hydroperoxide).

10.3.3 Add 100 μ L/well of ROS Orange Working Solution into the cell plate.

Δ Note: for 384 well plate, add 25 μ L/well.

10.3.4 Incubate cells in a 37°C/5% CO₂ incubator for 30 – 60 minutes.

10.3.5 Monitor fluorescence increase at Ex/Em = 540/570 nm (cut off = 550 nm) with bottom read mode.

11. Assay Procedure – Flow cytometry

- Equilibrate all materials and prepared reagents to room temperature prior to use.
- We recommend that you assay all controls and samples in duplicate.
- Prepare all reagents and samples as directed in the previous sections.

11.1 Grow cells:

11.1.1 Prepare cells at a density of $5 \times 10^5 - 10^6$ cells/mL.

Δ Note: Each cell line should be evaluated on an individual basis to determine the optimal cell density.

11.2 Run ROS assay:

11.2.1 Treat cells with test compounds in PBS or HHBS. For untreated cells, add equivalent amount of compound buffer.

11.2.2 Induce ROS production by incubating cell plate at room temperature or in a 37°C/5% CO₂ incubator for at least 15 minutes, or required time for your compound.

Δ Note: for example, incubate HeLa cells for 30 minutes when treated with 100 μM TBHP (tert-butyl-hydroperoxide).

11.2.3 Add 1 μL of ROS Orange Stock Solution (Step 9.3) to 1 mL of cells and mix well.

Δ Note: ROS Orange Stock Solution can be diluted 5-fold to 200X in DMSO for convenience. 200X stock solution can be aliquoted and stored as initial solution (Step 9.3). If using 200X Stock, add 5 μL of Orange Stain to 1 mL of cells.

11.2.4 Incubate cells in a 37°C/5% CO₂ incubator for 30 – 60 minutes.

11.2.5 Monitor fluorescence increase with a flow cytometer at Ex/Em = 540/570 nm (FL2 channel).

12. Data Analysis

FLUORESCENCE MICROPLATE MEASUREMENT

- Subtract blank readings from all measurements (control and treated)
- Using fluorescent intensity, determine fold change between control and treated cells.

FLOW CYTOMETRY MEASUREMENT

- Establish appropriate FSC vs SSC gates to exclude debris and cell aggregates.
- Using mean fluorescent intensity, determine fold change between control and treated cells.

13. Typical data

Data provided for **demonstration purposes** only.

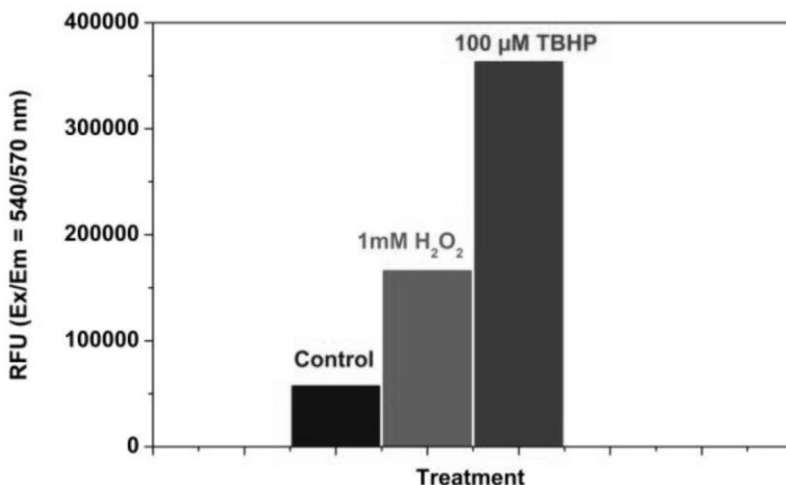


Figure 1. Detection of ROS in HeLa cells. HeLa cells were seeded overnight at 1.5×10^4 cells/90 μ L per well in a Costar black wall/clear bottom 96-well plate. Cells were untreated (control) or treated with 1 mM H₂O₂ or 100 μ M TBHP (ter-butyl hydroperoxide) for 30 minutes at 37°C. The ROS Orange stain solution (100 μ L/well) was added and cells were incubated in a 37°C/5% incubator for 1 hour. The fluorescence signal was monitored at Ex/Em = 540/570 nm (cut off 550 nm) with bottom read mode using FlexStation (Molecular Devices).

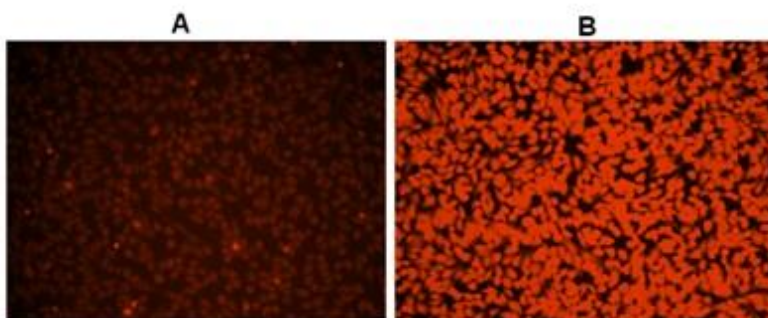


Figure 2. HeLa cells were stained with ROS Orange stain solution in a Costar black wall/clear bottom 96-well plate following protocol for microplate assay. A: Untreated control cells. B: Cells treated with 100 μ M TBHP (ter-butyl hydroperoxide) for 30 minutes before staining.

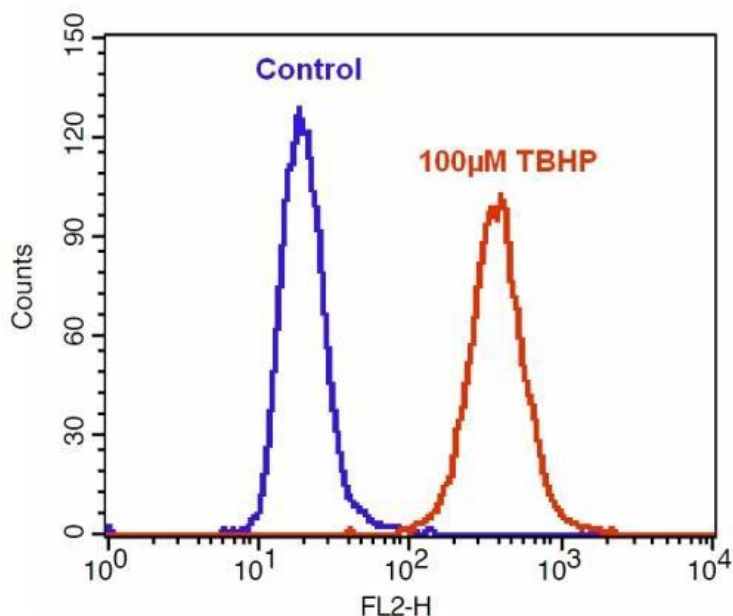


Figure 3. Detection of ROS in Jurkat cells. Jurkat cells were left untreated (blue) or treated with 100 μ M TBHP (ter-butyl hydroperoxide) (red) for 30 minutes at 37°C. The ROS Orange stain solution was added and cells were incubated in a 37°C/5% incubator for 1 hour. Fluorescent intensities were measured with a FACScalibur flow cytometer using FL2 channel.

14. FAQs

Q. Do any I to do any washing steps?

A. No washing steps are required for the staining procedure. However, we do recommend to change the culture media before addition of the test compounds.

Q. Can I fix my cells?

A. Cells can be fixed with a formaldehyde fixative after they have been stained with the ROS Orange dye. The signal will be retained after fixation.

15. Notes

Technical Support

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