ab189576 – Apolipoprotein A1 (APOA1) Human SimpleStep ELISA® Kit

Instructions for Use

For the quantitative measurement of Apolipoprotein A1 (APOA1) in human serum, plasma, and cell culture supernatants.

This product is for research use only and is not intended for diagnostic use.

Version 3 Last Updated 20 July 2023

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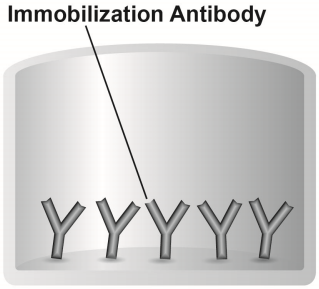
1. BACKGROUND

Apolipoprotein A1 (APOA1) *in vitro* SimpleStep ELISA® kit is designed for the quantitative measurement of Apolipoprotein A1 protein in human serum, plasma, and cell culture supernatants.

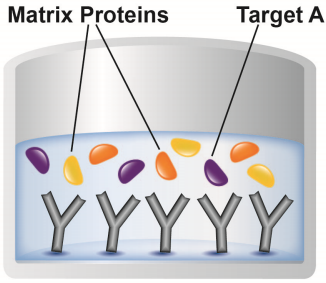
The SimpleStep ELISA® employs an affinity tag labeled capture antibody and a reporter conjugated detector antibody which immunocapture the sample analyte in solution. This entire complex (capture antibody/analyte/detector antibody) is in turn immobilized via immunoaffinity of an anti-tag antibody coating the well. To perform the assay, samples or standards are added to the wells, followed by the antibody mix. After incubation, the wells are washed to remove unbound material. TMB substrate is added and during incubation is catalyzed by HRP, generating blue coloration. This reaction is then stopped by addition of Stop Solution completing any color change from blue to yellow. Signal is generated proportionally to the amount of bound analyte and the intensity is measured at 450 nm. Optionally, instead of the endpoint reading, development of TMB can be recorded kinetically at 600 nm.

Apolipoprotein AI (ApoA-I) is secreted by the liver and small intestine and is a major protein of plasma HDL (high density lipoprotein). APOA-I participates in the reverse transport of cholesterol from tissues to the liver for excretion by promoting cholesterol efflux from tissues and by acting as a cofactor for the lecithin cholesterol acyltransferase (LCAT). Defects in APOA-I are associated with several diseases associated with low HDL levels (HDLD1 and HDLD2) and amyloidosis (AMYLIOWA and AMYL8).

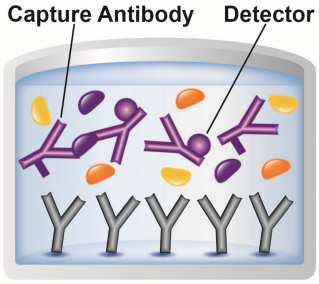
2. ASSAY SUMMARY



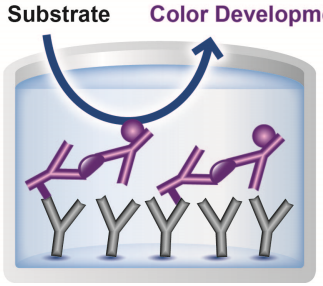
Remove appropriate number of antibody coated well strips. Equilibrate all reagents to room temperature. Prepare all reagents, samples, and standards as instructed.



Add standard or sample to appropriate wells.



Add Antibody Cocktail to all wells. Incubate at room temperature.



Aspirate and wash each well. Add TMB Substrate to each well and incubate. Add Stop Solution at a defined endpoint. Alternatively, record color development kinetically after TMB substrate addition.

3. PRECAUTIONS

Please read these instructions carefully prior to beginning the assay.

All kit components have been formulated and quality control tested to function successfully as a kit. Modifications to the kit components or procedures may result in loss of performance.

4. STORAGE AND STABILITY

Store kit at 2-8ºC immediately upon receipt.

Refer to list of materials supplied for storage conditions of individual components. Observe the storage conditions for individual prepared components in sections 9 & 10.

5. MATERIALS SUPPLIED

Item	Quantity 1 x 96 tests	Quantity 10 x 96 tests	Storage Condition (before preparation)
10X APOA1 Capture Antibody	600 µL	10 x 600 µL	+4ºC
10X APOA1 Detector Antibody	600 µL	10 x 600 µL	+4ºC
APOA1 Human Lyophilized Recombinant Protein	2 Vials	10 x 2 Vials	+4ºC
Antibody Diluent 5BI	6 mL	10 x 6 mL	+4ºC
Sample Diluent NS	50 mL	2 x 250 mL	+4ºC
Wash Buffer PT 10 X	20 mL	200 mL	+4ºC
TMB Development Solution	12 mL	120 mL	+4ºC
Stop Solution	12 mL	120 mL	+4ºC
Anti-tag coated microplate (12 x 8 well strips)	96 Wells	10 x 96 Wells	+4ºC
Plate Seal	1	10	+4ºC

6. MATERIALS REQUIRED, NOT SUPPLIED

These materials are not included in the kit, but will be required to successfully utilize this assay:

- Microplate reader capable of measuring absorbance at 450 or 600 nm.

- Method for determining protein concentration (BCA assay recommended).
- Deionized water
- PBS (1.4 mM KH2PO4, 8 mM Na2HPO4, 140 mM NaCl, 2.7 mM KCl, pH 7.4).
- Multi- and single-channel pipettes.
- Tubes for standard dilution.
- Plate shaker for all incubation steps.
- Optional: Phenylmethylsulfonyl Fluoride (PMSF) (or other protease inhibitors).

7. LIMITATIONS

- Assay kit intended for research use only. Not for use in diagnostic procedures.
- Do not mix or substitute reagents or materials from other kit lots or vendors. Kits are QC tested as a set of components and performance cannot be guaranteed if utilized separately or substituted.

8. TECHNICAL HINTS

- Samples generating values higher than the highest standard should be further diluted in the appropriate sample dilution buffers.
- Avoid foaming or bubbles when mixing or reconstituting components.
- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions.
- Ensure plates are properly sealed or covered during incubation steps.
- Complete removal of all solutions and buffers during wash steps is necessary to minimize background.

- As a guide, typical ranges of sample concentration for commonly used sample types are shown below in Sample Preparation (section 11).
- All samples should be mixed thoroughly and gently.
- Avoid multiple freeze/thaw of samples.
- Incubate ELISA plates on a plate shaker during all incubation steps
- When generating positive control samples, it is advisable to change pipette tips after each step.
- **To avoid high background always add samples or standards to the well before the addition of the antibody cocktail.**
- **This kit is sold based on number of tests. A 'test' simply refers to a single assay well. The number of wells that contain sample, control or standard will vary by product. Review the protocol completely to confirm this kit meets your requirements. Please contact our Technical Support staff with any questions.**

9. REAGENT PREPARATION

- Equilibrate all reagents to room temperature (18-25°C) prior to use. The kit contains enough reagents for 96 wells. **The sample volumes below are sufficient for 48 wells (6 x 8-well strips); adjust volumes as needed for the number of strips in your experiment.**
- Prepare only as much reagent as is needed on the day of the experiment. Capture and Detector Antibodies have only been tested for stability in the provided 10X formulations.

9.1 1X Wash Buffer PT

Prepare 1X Wash Buffer PT by diluting 10X Wash Buffer PT with deionized water. To make 50 mL 1X Wash Buffer PT combine 5 mL 10X Wash Buffer PT with 45 mL deionized water. Mix thoroughly and gently.

9.2 Antibody Cocktail

Prepare Antibody Cocktail by diluting the capture and detector antibodies in Antibody Diluent 5BI. To make 3 mL of the Antibody Cocktail combine 300 µL 10X Capture Antibody and 300 µL 10X Detector Antibody with 2.4 mL Antibody Diluent 5BI. Mix thoroughly and gently.

10. STANDARD PREPARATION

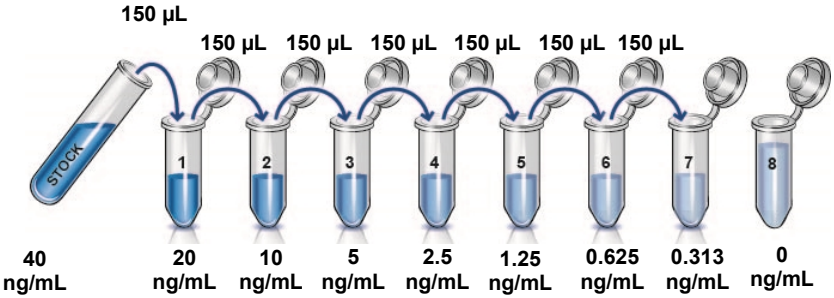
Prepare serially diluted standards immediately prior to use. Always prepare a fresh set of positive controls for every use.

The following section describes the preparation of a standard curve for duplicate measurements (recommended).

10.1 IMPORTANT: If the protein standard vial has a volume identified on the label, reconstitute the Apolipoprotein A1 standard by adding that volume of Sample Diluent NS indicated on the label. Alternatively, if the vial has a mass identified, reconstitute the Apolipoprotein A1 standard by adding 500 µL Sample Diluent NS. Hold at room temperature for 10 minutes and mix gently. This is the 40 ng/mL **Stock Standard Solution**.

10.2 Label eight tubes, Standards 1– 8 and add 150 µL Sample Diluent NS into each tube.

10.3 Use Standard #1 to prepare the following dilution series. Standard #8 contains no protein and is the Blank control:



11. SAMPLE PREPARATION

TYPICAL SAMPLE DYNAMIC RANGE	
Sample Type	**Range**
Human Serum	1:8x10^6 – 1: 2.5x10^5
Human Plasma - Citrate	1:8x10^6 – 1: 2.5x10^5
Human Plasma - EDTA	1:8x10^6 – 1: 2.5x10^5
Human Plasma - Heparin	1:8x10^6 – 1: 2.5x10^5

11.1 Plasma

Collect plasma using citrate, EDTA or heparin. Centrifuge samples at 2,000 x g for 10 minutes. Dilute samples into Sample Diluent NS and assay. Store un-diluted plasma samples at -20ºC or below for up to 3 months. Avoid repeated freeze-thaw cycles.

11.2 Serum

Samples should be collected into a serum separator tube. After clot formation, centrifuge samples at 2,000 x g for 10 minutes and collect serum. Dilute samples into Sample Diluent NS and assay. Store un-diluted serum at -20ºC or below. Avoid repeated freeze-thaw cycles.

11.3 Cell Culture Supernatants

Centrifuge cell culture media at 2,000 x g for 10 minutes to remove debris. Collect supernatants and assay. Store samples at -20°C or below. Avoid repeated freeze-thaw cycles.

Note: Due to the high dilutions required for certain Human serum and plasma samples, we recommend initially diluting your samples in 1X Wash Buffer and then performing the final dilution in Sample Diluent NS. As an example the table below demonstrates the steps suggested to generate a final sample dilution of $1:2.5x10^5$:

Tube #	Sample to Dilute	Volume of Sample (µL)	Volume of 1X Wash Buffer (µL)	Volume of Sample Diluent NS (µL)	Starting Conc.	Final Conc.
1	Neat serum/ plasma	6	594	-	Neat	1:100
2	Tube #1	6	594	-	1:100	1:10,000
3	Tube #2	12	-	288	1:10,000	$1:2.5x10^5$

12. PLATE PREPARATION

- The 96 well plate strips included with this kit are supplied ready to use. It is not necessary to rinse the plate prior to adding reagents
- Unused plate strips should be immediately returned to the foil pouch containing the desiccant pack, resealed and stored at 4°C
- For each assay performed, a minimum of two wells must be used as the zero control
- For statistical reasons, we recommend each sample should be assayed with a minimum of two replicates (duplicates)
- Differences in well absorbance or “edge effects” have not been observed with this assay

13. ASSAY PROCEDURE

- **Equilibrate all materials and prepared reagents to room temperature prior to use.**
- **It is recommended to assay all standards, controls and samples in duplicate.**

13.1 Prepare all reagents, working standards, and samples as directed in the previous sections.

13.2 Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, reseal and return to 4ºC storage.

13.3 Add 50 µL of all sample or standard to appropriate wells.

13.4 Add 50 µL of the Antibody Cocktail to each well.

13.5 Seal the plate and incubate for 1 hour at room temperature on a plate shaker set to 400 rpm.

13.6 Wash each well with 3 x 350 µL 1X Wash Buffer PT. Wash by aspirating or decanting from wells then dispensing 350 µL 1X Wash Buffer PT into each well. Complete removal of liquid at each step is essential for good performance. After the last wash invert the plate and blot it against clean paper towels to remove excess liquid.

13.7 Add 100 µL of TMB Substrate to each well and incubate for 10 minutes in the dark on a plate shaker set to 400 rpm.

Given variability in laboratory environmental conditions, optimal incubation time may vary between 5 and 20 minutes. Note: The addition of Stop Solution will change the color from blue to yellow and enhance the signal intensity about 3X. To avoid signal saturation, proceed to the next step before the high concentration of the standard reaches a blue color of O.D.600 equal to 1.0.

13.8 Add 100 µL of Stop Solution to each well. Shake plate on a plate shaker for 1 minute to mix. Record the OD at 450 nm. This is an endpoint reading.

Alternative to 13.7 – 13.8: Instead of the endpoint reading at 450 nm, record the development of TMB Substrate kinetically. Immediately after addition of TMB Development Solution begin recording the blue color development with elapsed time in the microplate reader prepared with the following settings:

Mode:	Kinetic
Wavelength:	600 nm
Time:	up to 20 min
Interval:	20 sec - 1 min
Shaking:	Shake between readings

Note that an endpoint reading can also be recorded at the completion of the kinetic read by adding 100 µL Stop Solution to each well and recording the OD at 450 nm.

13.9 Analyze the data as described below.

14. CALCULATIONS

14.1 Calculate the average absorbance value for the blank control (zero) standards. Subtract the average blank control standard absorbance value from all other absorbance values.

14.2 Create a standard curve by plotting the average blank control subtracted absorbance value for each standard concentration (y-axis) against the target protein concentration (x-axis) of the standard. Use graphing software to draw the best smooth curve through these points to construct the standard curve.

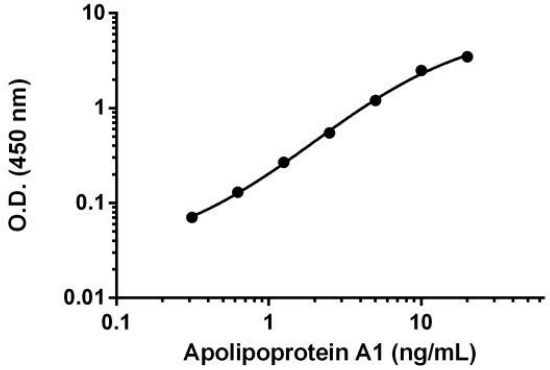
Note: Most microplate reader software or graphing software will plot these values and fit a curve to the data. A four parameter curve fit (4PL) is often the best choice; however, other algorithms (e.g. linear, semi-log, log/log, 4 parameter logistic) can also be tested to determine if it provides a better curve fit to the standard values.

14.3 Determine the concentration of the target protein in the sample by interpolating the blank control subtracted absorbance values against the standard curve. Multiply the resulting value by the appropriate sample dilution factor, if used, to obtain the concentration of target protein in the sample.

14.4 Samples generating absorbance values greater than that of the highest standard should be further diluted and reanalyzed. Similarly, samples which measure at an absorbance values less than that of the lowest standard should be retested in a less dilute form.

15. TYPICAL DATA

TYPICAL STANDARD CURVE – Data provided for **demonstration purposes only**. A new standard curve must be generated for each assay performed.



Standard Curve Measurements			
Conc. (ng/mL)	O.D. 450 nm		Mean O.D.
	1	2	
0	0.06	0.06	0.06
0.313	0.12	0.11	0.11
0.625	0.16	0.17	0.16
1.25	0.30	0.30	0.30
2.5	0.60	0.57	0.58
5.0	1.23	1.19	1.21
10	2.60	2.34	2.48
20	3.62	3.62	3.62

Figure 1. Example of Apolipoprotein A1 standard curve. The Apolipoprotein A1 standard curve was prepared as described in Section 10. Raw data values are shown in the table. Background-subtracted data values (mean +/- SD) are graphed.

16. TYPICAL SAMPLE VALUES

SENSITIVITY –

The calculated minimal detectable dose (MDD) is 59 pg/mL with a range of 24-93ng/mL The MDD was determined by calculating the mean of zero standard replicates (n=56) and adding 2 standard deviations then extrapolating the corresponding concentrations.

RECOVERY –

Three concentrations of Apolipoprotein A1 were spiked in duplicate to the indicated biological matrix to evaluate signal recovery in the working range of the assay.

Sample Type	Average % Recovery	Range (%)
Human Serum	115	111-122
Human Plasma - Citrate	119	117-123
Human Plasma - EDTA	112	108-115
Human Plasma - Heparin	95	76-111
Cell Culture Media	89	85-92

LINEARITY OF DILUTION –

Native Apolipoprotein A1 was measured in Human serum and plasma samples in a 2-fold dilution series. Sample dilutions were made in Sample Diluent NS.

Recombinant Apolipoprotein A1 was spiked into cell culture media and diluted in a 2-fold dilution series in Sample Diluent NS.

Linearity of dilution is determined based on interpolated values from the standard curve. Linearity of dilution defines a sample concentration interval in which interpolated target concentrations are directly proportional to sample dilution.

Dilution Factor	Interpolated value	0.0002% Human Plasma (Citrate)	0.0002% Human Plasma (EDTA)	0.0002% Human Plasma (Heparin)
Undiluted	ng/mL	2.79	3.88	3.12
	% Expected value	**100**	**100**	**100**
2	ng/mL	1.41	1.81	1.52
	% Expected value	**101**	**93**	**98**
4	ng/mL	0.72	0.94	0.75
	% Expected value	**103**	**97**	**97**
8	ng/mL	0.36	0.46	0.37
	% Expected value	**103**	**94**	**96**
16	ng/mL	**0.17**	**0.23**	**0.18**
	% Expected value	**98**	**93**	**93**

Dilution Factor	Interpolated value	0.0002% Human Serum	25% Cell culture supernatant
Undiluted	ng/mL	3.09	4.09
	% Expected value	**100**	**100**
2	ng/mL	1.50	2.22
	% Expected value	**97**	**108**
4	ng/mL	0.77	1.15
	% Expected value	**100**	**113**
8	ng/mL	0.38	0.59
	% Expected value	**97**	**115**
16	ng/mL	0.18	**0.28**
	% Expected value	**92**	**111**

PRECISION –

Mean coefficient of variations of interpolated values from 3 concentrations of Apolipoprotein A1 within the working range of the assay.

	Intra-Assay	Inter-Assay
n=	5	3
CV (%)	2.1	2.9

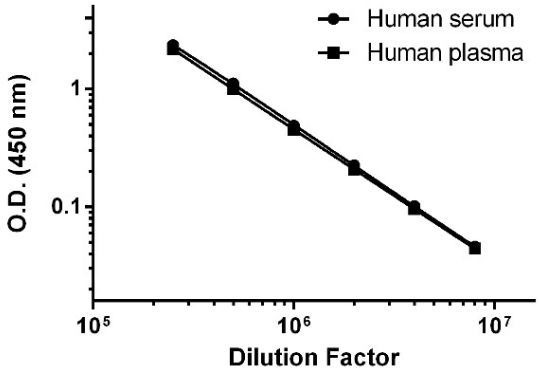


Figure 2. Titration of human serum and plasma (citrate) diluted 2.5x10^5 –fold to 8x10^6 –fold in Sample Diluent NS. Background subtracted data from duplicate measurements are plotted.

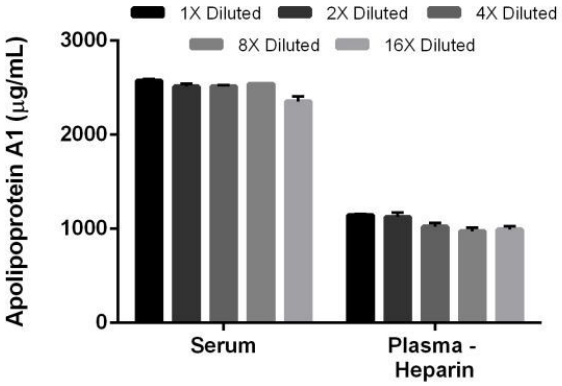


Figure 3. Interpolated concentrations of Apolipoprotein A1 in human serum and plasma heparin. The concentrations of Apolipoprotein A1 were measured in duplicate and interpolated from the Apolipoprotein A1 standard curve and corrected for sample dilution. The interpolated dilution factor corrected values are plotted (mean +/- SD, n=2). The mean Apolipoprotein A1 concentration was determined to be 2,500 µg/mL in serum and 1,053ug/mL in plasma heparin.

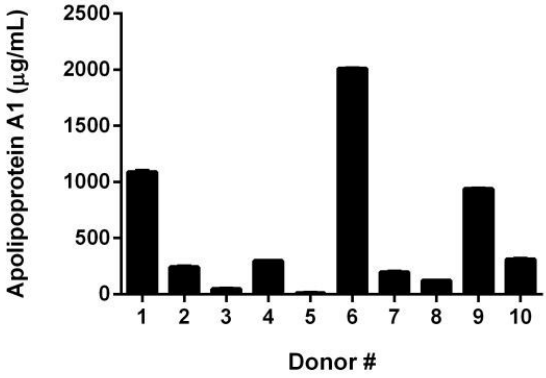


Figure 4. Interpolated concentrations of Apolipoprotein A1 in human serum from 10 donors. Serum from 10 apparently healthy male donors was measured in duplicate. The mean Apolipoprotein A1 concentration was determined to be 526 µg/mL with a range of 9.8-2,011 µg/mL.

17. ASSAY SPECIFICITY

This kit recognizes both native and recombinant human Apolipoprotein A1 protein in serum, plasma and cell culture supernatant samples only.

Cell and tissue extract samples have not been tested with this kit.

CROSS REACTIVITY -

The proteins listed below were prepared at 50 ng/mL in Sample Diluent NS and assayed for cross-reactivity. No significant cross-reactivity was observed.

Recombinant human:

Apolipoprotein B (APOB)

INTERFERENCE -

The recombinant human Apolipoprotein B was prepared at 50 ng/mL in Sample Diluent NS in the presence of 250 pg/mL Apolipoprotein A1 and assayed for interference. No interference was observed.

18. SPECIES REACTIVITY

This kit recognizes human Apolipoprotein A1 protein.

Other species reactivity was determined by measuring 1:1.25x10^5 dilution of serum samples of various species, interpolating the protein concentrations from the human standard curve, and expressing the interpolated concentrations as a percentage of the protein concentration in human serum assayed at the same dilution.

Reactivity < 3% was determined for the following species:

- Mouse
- Rat
- Hamster
- Guinea Pig
- Rabbit
- Dog
- Goat
- Pig
- Cow
- Goat
- Chicken

Please contact our Technical Support team for more information

19. TROUBLESHOOTING

Problem	Cause	Solution
Difficulty pipetting lysate; viscous lysate.	Genomic DNA solubilized	Prepare 1X Cell Extraction Buffer PTR (without enhancer). Add enhancer to lysate after extraction.
Poor standard curve	Inaccurate Pipetting	Check pipettes
	Improper standard dilution	Prior to opening, briefly spin the stock standard tube and dissolve the powder thoroughly by gentle mixing
Low Signal	Incubation times too brief	Ensure sufficient incubation times; increase to 2 or 3 hour standard/sample incubation
	Inadequate reagent volumes or improper dilution	Check pipettes and ensure correct preparation
	Incubation times with TMB too brief	Ensure sufficient incubation time until blue color develops prior addition of Stop solution
Large CV	Plate is insufficiently washed	Review manual for proper wash technique. If using a plate washer, check all ports for obstructions.
	Contaminated wash buffer	Prepare fresh wash buffer
Low sensitivity	Improper storage of the ELISA kit	Store your reconstituted standards at -80°C, all other assay components 4°C. Keep TMB substrate solution protected from light.
Precipitate in Diluent	Precipitation and/or coagulation of components within the Diluent.	Precipitate can be removed by gently warming the Diluent to 37ºC.

20. **NOTES**

For all technical or commercial enquiries please go to:

www.abcam.com/contactus
www.abcam.cn/contactus (China)
www.abcam.co.jp/contactus (Japan)