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ab237645

Anti-HER2 ELISA Kit

For the measurement of anti-HER2 in human serum and plasma.

[View kit datasheet: www.abcam.com/ab237645](http://www.abcam.com/ab237645)

(use www.abcam.cn/ab237645 for China, or www.abcam.co.jp/ab237645 for Japan)

This product is for research use only and is not intended for diagnostic use.

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1. Overview

Anti-HER2 ELISA Kit (ab237645) is a highly specific and sensitive kit designed for the in vitro determination of anti-HER2 in biological matrices such as human serum and plasma. The density of color is proportional to the amount of anti-HER2 captured from the samples and can be quantified when compared with standard curve.

This product is a recombinant DNA-derived humanized monoclonal antibody that selectively targets the extracellular domain of the human epidermal growth factor receptor 2 protein (HER2). Anti-HER2 has anti-tumor activity against HER2-positive human breast tumor cells in laboratory models and is active for the treatment of women with HER2-overexpressing breast cancers. This antibody was approved in 1998 for clinical use for HER2 overexpressing metastatic breast cancer. In HER2 overexpressing cells, anti-HER2 markedly down-regulates HER2 expression by accelerating receptor endocytosis and degradation and inhibits cell cycle progression by inducing the formation of p27Kip1/Cdk2 complexes.

2. Protocol Summary

Prepare all reagents, samples, and standards as instructed



Add 100 μ L of standard or sample to appropriate wells. Cover and incubate for 30 minutes at room temperature



Discard incubation solution and wash plate 3 times with 300 μ L diluted Wash Buffer



Add 100 μ L HRP-conjugate to each well. Cover and incubate for 30 minutes at room temperature



Discard the solution and wash plate 3 times with 300 μ L diluted Wash Buffer



Add 100 μ L TMB Substrate and incubate the plate in the dark at room temperature for 10 minutes.



Add 100 μ L Stop Solution and read OD at 450 nm within 20 minutes.

3. Precautions

Please read these instructions carefully prior to beginning the assay.

- Observe good laboratory practices. Gloves, lab coat, and protective eyewear should always be worn. Never pipet by mouth. Do not eat, drink or smoke in the laboratory areas.
- For general guidelines, precautions, limitations on the use of our assay kits and general assay troubleshooting tips, particularly for first time users, please consult our guide: <https://www.abcam.com/en-us/products/biochemical-assays>
- If applicable, please refer to the current Safety Data Sheet (SDS) provided with this product for safety, handling, and disposal information. The most up to date and current versions are available on our website <https://www.abcam.com/en-us>.

4. Storage and Stability

Store kit at +4°C immediately upon receipt.

Refer to list of materials supplied for storage conditions of individual components.

5. Limitations

- Assay kit intended for research use only. Not for use in diagnostic procedures.
- Do not mix or substitute reagents or materials from other kit lots or vendors.

6. Materials Supplied

Item	Quantity	Storage condition
Micro ELISA Plate	1 unit	+4°C
Anti-HER2 Standard S1	300 µL	+4°C
Anti-HER2 Standard S2	300 µL	+4°C
Anti-HER2 Standard S3	300 µL	+4°C
Anti-HER2 Standard S4	300 µL	+4°C
Anti-HER2 Standard S5	300 µL	+4°C
Anti-HER2 Standard S6	300 µL	+4°C
Anti-HER2 Standard S7	300 µL	+4°C
Assay Buffer	2 x 50 mL	+4°C
HRP-conjugate Probe	12 mL	+4°C
TMB Substrate	12 mL	+4°C
Stop Solution	12 mL	+4°C
Wash Buffer (20X)	50 mL	+4°C
Plate sealers	2 units	+4°C

7. Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully perform this assay:

- Microplate reader capable of measuring absorbance at OD 450 nm
 - Deionized water.
 - Multi- and single-channel pipettes.
 - Tubes for sample dilution.
 - Plate shaker for all incubation steps.
 - Absorbent paper

8. Technical Hints

- Samples generating values higher than the highest standard should be further diluted.
- Avoid foaming or bubbles when mixing or reconstituting components.
- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions.
- Ensure plates are properly sealed or covered during incubation steps.
- Complete removal of all solutions and buffers during wash steps is necessary to minimize background.
- All samples should be mixed thoroughly and gently.
- Avoid multiple freeze/thaw of samples.
- Incubate ELISA plates on a plate shaker during all incubation steps.
- When generating positive control samples, it is advisable to change pipette tips after each step.

9. Reagent Preparation

- Equilibrate all reagents to room temperature (18-25°C) prior to use. Before using the kit, spin tubes and bring down all components to the bottom of tubes.
- Prepare only as much reagent as is needed on the day of the experiment.

9.1 20X Wash Buffer:

Dilute the 20X Wash Buffer to 1X solution in ddH₂O (10 mL of Wash Buffer stock to 190 mL of ddH₂O). Mix the 1X solution thoroughly by vortex manually. The working stock can be stable for 2 weeks after preparation at 4°C.

10. Standard and Control Preparation

- Always prepare a fresh set of standards for every use.
- Discard working standard dilutions after use as they do not store well.

Dilute the 10X stock solutions of standards with Assay buffer (20 µL standards to 180 µL Assay buffer). Please see table below for concentrations:

Name	S1	S2	S3	S4	S5	S6	S7
Stock conc. ng/mL	3000	1000	300	100	0	High control	Low control
Working conc. ng/mL	300	100	30	10	0	-	-

11. Sample Preparation

General sample information:

- We recommend performing several dilutions of your sample to ensure the readings are within the standard value range.
- We recommend that you use fresh samples for the most reproducible assay.

11.1 Serum/plasma:

- 11.1.1 Dilute samples at 1:10 (10 µL serum/plasma + 90 µL Assay buffer), then dilute another 1:100 (5 µL diluted serum/plasma + 495 µL Assay buffer) to a total of 1:1000 dilution
- 11.1.2 Diluted samples should further be diluted if the concentration of anti-HER2 is higher than the measuring range.
- 11.1.3 The usual precautions for venipuncture should be observed. Samples are stable at 4°C for 2 days and -20°C for 6 months. Avoid freeze-and-thaw cycle.

Δ Note: The usual precautions for venipuncture should be observed.

12. Assay Procedure

- Prepare reagents within 30 minutes before the experiment.
 - Equilibrate all materials and prepared reagents to room temperature 15 minutes prior to use.
 - We recommend that you assay all standards, controls and samples in duplicate.
-
- 12.1** Add 100 μ L of standards and diluted samples into appropriate wells. Cover wells and incubate for 30 minutes at room temperature.
 - 12.2** Discard incubation solution. Wash plate 3 times each with 300 μ L of diluted Wash Buffer. Remove excess solution by tapping the inverted plate on a paper towel.
 - 12.3** Add 100 μ L of HRP-conjugate into each well. Cover wells with adhesive plate sealer and incubate at room temperature for 30 minutes.
 - 12.4** Discard the solution and wash the wells as step 12.2.
 - 12.5** Add 100 μ L of 1X TMB substrate solution and incubate the plate in the dark at room temperature for 10 minutes.
 - 12.6** Add 100 μ L of Stop solution to stop the reaction.
 - 12.7** Read the absorbance in a microplate reader set to 450 nm within 20 minutes. (Reference wavelength to 650 nm).

13. Calculations

- 13.1 Calculate the average absorbance value for the blank control (0 ng/mL) standards. Subtract the average blank control standard absorbance value from all other absorbance values.
- 13.2 **Create a standard curve** by plotting the average blank control subtracted absorbance value for each standard concentration (y axis) against the target protein concentration (x axis) of the standard.
- 13.3 Construct a standard curve of difference data using software capable of generating four-parameter logistic (4PL) or point-to-point calculation curve fit.
- 13.4 To obtain the exact values of the samples, the concentration determined from the standard curve should be multiplied by the dilution factor.
- 13.5 Determine the concentration of the target protein in the sample by interpolating the blank control subtracted **absorbance values against the standard curve**. Multiply the resulting value by the appropriate dilution factor to obtain the concentration of target protein in the sample.
- 13.6 Samples generating absorbance values greater than that of the highest standard should be further diluted and reanalyzed. Similarly, samples which measure at an absorbance values less than that of the lowest standard should be retested in a less dilute form.

14. Typical Data

Typical standard curve - data provided **for demonstration purposes only**. A new standard curve must be generated for each assay performed.

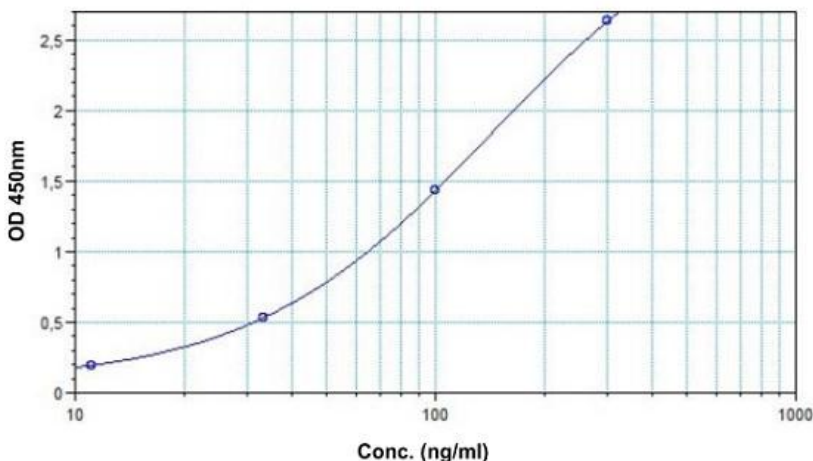


Figure 1. Typical Standard Curve: This standard curve is for demonstration only. A standard curve must be run with each assay.

Typical Sample Values

- **Detection Range:** 10 - 300 ng/ml
- **Sensitivity:** Quantitative limit - 10 ng/ml, Detection limit - 2 ng/ml
- **Assay Precision:** Intra-Assay: CV < 30%; Inter-Assay: CV < 30% (CV (%) = SD/mean X 100)
- **Recovery rate:** <100±30% with normal human serum samples with known concentrations
- **Cross Reactivity:** No significant cross-reactivity or interference with other proteins present in native human serum or other therapeutic immunoglobulins.

15. Notes

Technical Support

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