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ab237655 anti-HER2 mab ELISA Kit

For the measurement of the antibody against anti-HER2 mab in human serum and plasma.

This product is for research use only and is not intended for diagnostic use.

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1. Overview

Anti-HER2 mab ELISA Kit (ab237655) is designed to quantify/measure the antibody against anti-HER2 mab with high specificity and sensitivity in biological matrices.

Anti-HER2 mab(Trastuzumab) is a recombinant DNA-derived humanized monoclonal antibody that selectively targets the extracellular domain of the human epidermal growth factor receptor 2 protein (HER2). It has antitumor activity against HER2-positive human breast tumor cells in laboratory models and is active for the treatment of women with HER2-overexpressing breast cancers. In HER2 overexpressing cells, this antibody markedly down-regulates HER2 expression by accelerating receptor endocytosis and degradation and inhibits cell cycle progression by inducing the formation of p27Kip1/Cdk2 complexes. However, some patients develop unwanted immunogenicity, which leads to production of anti-drug-antibodies (ADAs) inactivating the therapeutic effects of the treatment and, in rare cases, inducing adverse effects.

2. Protocol Summary

Prepare all reagents, samples, and standards as instructed

Add 100 µL standard, controls, diluted sample and confirmation test mixture to appropriate wells. Cover and incubate for 60 minutes at room temperature

Discard incubation solution and wash plate 3 times with 300 µL diluted Wash Buffer

Add 100 µL peroxidase conjugate to each well. Cover and incubate for 60 minutes at room temperature

Discard the solution and wash plate 3 times with 300 µL diluted Wash Buffer

Add 100 µL TMB Substrate and incubate the plate in the dark at room temperature for 20 minutes.

Add 100 µL Stop Solution and read OD at 450 nm within 20 minutes.

3. Precautions

Please read these instructions carefully prior to beginning the assay.

- Reagents should be treated as possible mutagens and should be handle with care and disposed of properly. Please review the Safety Datasheet (SDS) provided with the product for information on the specific components.
- Observe good laboratory practices. Gloves, lab coat, and protective eyewear should always be worn. Never pipet by mouth. Do not eat, drink or smoke in the laboratory areas.
- For general guidelines, precautions, limitations on the use of our assay kits and general assay troubleshooting tips, particularly for first time users, please consult our guide: www.abcam.com/assaykitguidelines
- All biological materials should be treated as potentially hazardous and handled as such. They should be disposed of in accordance with established safety procedures.

4. Storage and Stability

Store kit at +4°C immediately upon receipt. Kit has a storage time of 1 year from receipt, providing components have not been reconstituted.

Refer to list of materials supplied for storage conditions of individual components.

5. Limitations

- Assay kit intended for research use only. Not for use in diagnostic procedures.
- Do not mix or substitute reagents or materials from other kit lots or vendors.

6. Materials Supplied

Item	Quantity	Storage

		condition
Micro ELISA Plate	1 unit	+4°C
Anti-HER2 mab Standard S1	1 mL	+4°C
Anti-HER2 mab Standard S2	1 mL	+4°C
Anti-HER2 mab Standard S3	1 mL	+4°C
Anti-HER2 mab Standard S4	1 mL	+4°C
Anti-HER2 mab Standard S5	1 mL	+4°C
Anti-HER2 mab Standard S6	1 mL	+4°C
Anti-HER2 mab Standard S7	1 mL	+4°C
Confirmation Reagent	12 mL	+4°C
Assay Buffer	50 mL	+4°C
Peroxidase Conjugate	12 mL	+4°C
TMB Substrate	12 mL	+4°C
Stop Solution	12 mL	+4°C
Wash Buffer (20X)	50 mL	+4°C
Plate sealers	2 units	+4°C

7. Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully perform this assay:

- Microplate reader capable of measuring absorbance at OD 450 nm
- Deionized water.
- Multi- and single-channel pipettes.
- Tubes for sample dilution.
- Plate shaker for all incubation steps.
- Absorbent paper

8. Technical Hints

- Samples generating values higher than the highest standard should be further diluted.
- Avoid foaming or bubbles when mixing or reconstituting components.
- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions.
- Ensure plates are properly sealed or covered during incubation steps.
- Complete removal of all solutions and buffers during wash steps is necessary to minimize background.
- All samples should be mixed thoroughly and gently.
- Avoid multiple freeze/thaw of samples.
- Incubate ELISA plates on a plate shaker during all incubation steps.
- When generating positive control samples, it is advisable to change pipette tips after each step.

9. Reagent Preparation

- Equilibrate all reagents to room temperature (18-25°C) prior to use. Before using the kit, spin tubes and bring down all components to the bottom of tubes.
- Prepare only as much reagent as is needed on the day of the experiment.

9.1 20X Wash Buffer:

Dilute the 20X Wash Buffer to 1X solution in ddH_2O (10 mL of Wash Buffer stock to 190 mL of ddH_2O). Mix the 1X solution thoroughly by vortex manually. The working stock can be stable for 2 weeks after preparation at 4°C.

10. Standard and Control Preparation

Standard and controls, S1 – S7, are ready to use, please see table below for concentrations:

Name	S1	S2	S3	S4	S5	S6	S7
Conc. ng/mL	500	250	125	62	0	High control	Low control

Concentration for high and low controls are indicated on vials.

11. Sample Preparation

General sample information:

- We recommend performing several dilutions of your sample to ensure the readings are within the standard value range.
- We recommend that you use fresh samples for the most reproducible assay.

11.1 Serum/plasma:

1. Dilute samples at 1:10 (20 µL serum/plasma + 180 µL Assay buffer) or 1:100 (5 µL serum/plasma + 495 µL ddH_2O).
2. Diluted samples should further be diluted if the concentration of anti-HER2 mab is higher than the measuring range.
3. Samples are stable at 4°C for 7 days and -20°C for 6 months. Avoid freeze-and-thaw cycle.

Δ Note: The usual precautions for venipuncture should be observed.

12. Confirmation test mixture preparation

Mix 20 µL undiluted (positive) serum/plasma sample with 180 µL of confirmation reagent for 60 minutes in a microtube prior to the test.

13. Assay Procedure

- Prepare reagents within 30 minutes before the experiment.
- Equilibrate all materials and prepared reagents to room temperature 15 minutes prior to use.
- We recommend that you assay all standards, controls and samples in duplicate.

13.1 Add 100 µL of standards, controls, diluted samples and confirmation test mixture into appropriate wells. Cover wells and incubate for 60 minutes at room temperature.

13.2 Discard incubation solution. Wash plate 3 times each with 300 µL of diluted Wash Buffer. Remove excess solution by tapping the inverted plate on a paper towel.

13.3 Add 100 µL of Peroxidase Conjugate into each well. Cover wells with adhesive plate sealer and incubate at room temperature for 60 minutes.

13.4 Discard the solution and wash the wells as step 13.2.

13.5 Add 100 µL of 1X TMB substrate solution and incubate the plate in the dark at room temperature for 20 minutes.

13.6 Add 100 µL of Stop solution to stop the reaction.

13.7 Read the absorbance in a microplate reader set to 450 nm within 20 minutes. (Reference wavelength to 650 nm).

14. Calculations

14.1 Quantitative Calculation

14.1.1 Calculate the average absorbance value for the blank control (0 ng/mL) standards. Subtract the average blank control standard absorbance value from all other absorbance values.

14.1.2 Create a standard curve by plotting the average blank control subtracted absorbance value for each standard concentration (y axis) against the target protein concentration (x axis) of the standard.

14.1.3 Construct a standard curve of difference data using software capable of generating four-parameter logistic (4PL) or point-to-point calculation curve fit.

14.1.4 To obtain the exact values of the samples, the concentration determined from the standard curve should be multiplied by the dilution factor.

14.1.5 Determine the concentration of the target protein in the sample by interpolating the blank control subtracted absorbance values against the standard curve. Multiply the resulting value by the appropriate dilution factor to obtain the concentration of target protein in the sample.

14.1.6 Samples generating absorbance values greater than that of the highest standard should be further diluted and reanalyzed. Similarly, samples which measure at an absorbance values less than that of the lowest standard should be retested in a less dilute form.

14.2 Qualitative Interpretation

14.2.1 If "Sample OD450/650 / Zero Standard (S5) OD450/650" is < 3, the sample is NEGATIVE for antibody to anti-HER2 mab.

14.2.2 If "Sample OD450/650 /Zero Standard (S5) OD450/650" is ≥3, the sample is POSITIVE for antibody to anti-HER2 mab, and if required samples may be extrapolated for quantitative analysis and confirmation.

14.2.3 For the run to be valid, the OD450/650 nm of High Control should be ≥ 1.000 and the OD450/650 nm of each Low Control should be <0.200, if not, improper technique or reagent deterioration may be suspected and the run should be repeated.

14.2.4 Interpretation of true and false positive: For true positive sample, inhibition should be equal or greater than 25%.

$$\frac{OD\ sample - OD\ sample\ with\ confirmation\ reagent}{OD\ sample}\ x\ 100 = inhibition\ \%$$

15. Typical Data

Typical standard curve - data provided **for demonstration purposes only**. A new standard curve must be generated for each assay performed.

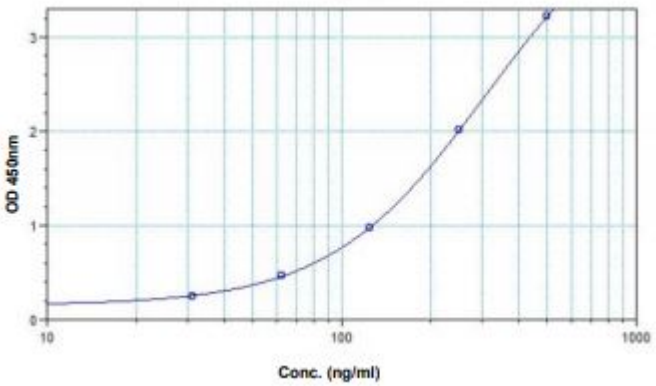

Figure 1. Typical Standard Curve: This standard curve is for demonstration only. A standard curve must be run with each assay.

16. Typical Sample Values

Detection Range: 62 - 500 ng/mL.

Sensitivity: 30 ng/mL.

Assay Precision: Intra-Assay: CV < 15%; Inter-Assay: CV < 15%

(CV (%) = SD/mean X 100)

Cross Reactivity: Anti-HER2 mab (Trastuzumab) infusion camouflages/masks the presence of antibody to Anti-HER2 mab in serum/plasma samples. Therefore, blood sampling time is critical for detection of antibody to Anti-HER2 mab. It is convenient to obtain blood sample just before the infusion of Anti-HER2 mab or at least 2 weeks after the infusion of Anti-HER2 mab.

Recovery rate: 85 – 115% with normal human serum samples with known concentrations.

17. Notes

Technical Support

For all technical or commercial enquiries please go to:

www.abcam.com/contactus

www.abcam.cn/contactus (China)

www.abcam.co.jp/contactus (Japan)