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ab272780
Rat Prolactin ELISA kit

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For the quantitative determination of prolactin antigen in rat plasma.

This product is for research use only and is not intended for diagnostic use.

Table of Contents

1. Overview

Rat Prolactin ELISA Kit is intended for the quantitative determination of prolactin antigen in rat plasma.

Rat prolactin will bind to the affinity purified capture antibody coated on the microtiter plate. After appropriate washing steps, anti‑rat prolactin primary antibody binds to the captured protein. Excess primary antibody is washed away and bound antibody is reacted with anti-rabbit HRP secondary reagent. Following an additional washing step, TMB substrate is used for color development which is measured at 450nm. A standard calibration curve is prepared along with the samples to be measured using dilutions of rat prolactin. Color development is proportional to the concentration of prolactin in the samples.

2. Protocol Summary

Prepare all reagents, samples, and standards as instructed.

Add 100 µL standard or sample to appropriate wells and shake plate at 300 rpm for 30 mins. Wash wells three times with 300 µL wash buffer. Remove excess wash by gently tapping plate on paper towel.

Add 100 µL of primary antibody to each well and shake plate at 300 rpm for 30 mins. Wash wells three times with 300 µL wash buffer. Remove excess wash by gently tapping plate on paper towel.

Add 100 µL of HRP secondary reagent to each well and shake plate at 300 rpm for 30 mins. Wash wells three times with 300 µL wash buffer. Remove excess wash by gently tapping plate on paper towel.

Add 100 µL TMB Substrate Solution to each well and shake plate for 2-10 mins.

Stop reaction with 50 µL H_2SO_4 or HCl stop solution, then measure the absorbance in all wells at 450nm.

3. Precautions

Please read these instructions carefully prior to beginning the assay.

- All kit components have been formulated and quality control tested to function successfully as a kit.
- We understand that, occasionally, experimental protocols might need to be modified to meet unique experimental circumstances. However, we cannot guarantee the performance of the product outside the conditions detailed in this protocol booklet.
- Reagents should be treated as possible mutagens and should be handle with care and disposed of properly. Please review the Safety Datasheet (SDS) provided with the product for information on the specific components.
- Observe good laboratory practices. Gloves, lab coat, and protective eyewear should always be worn. Never pipet by mouth. Do not eat, drink or smoke in the laboratory areas.
- All biological materials should be treated as potentially hazardous and handled as such. They should be disposed of in accordance with established safety procedures.

4. Storage and Stability

Store kit at +4°C immediately upon receipt. Kit has a storage time of 1 year from receipt, providing components have not been reconstituted.

Refer to list of materials supplied for storage conditions of individual components.

5. Limitations

- Assay kit intended for research use only. Not for use in diagnostic procedures.
- Do not mix or substitute reagents or materials from other kit lots or vendors. Kits are QC tested as a set of components and performance cannot be guaranteed if utilized separately or substituted.

6. Materials Supplied

Item	Quantity	Storage Condition
Rat Prolactin ELISA Plate	1 x 96 tests	+4°C
10X Wash Buffer	1 x 50 mL	+4°C
Rat Prolactin Standard Lyophilized Vial	1 vial	+4°C
Anti-Rat Prolactin Primary Antibody Lyophilized Vial	1 vial	+4°C
Anti-Rabbit HRP Secondary Reagent	1 vial	+4°C
TMB Substrate	1 x 10 mL	+4°C

7. Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully perform this assay:

- Microplate plate shaker capable of 300 rpm uniform horizontal circular movement
- Manifold dispenser/aspirator or automated microplate washer
- Microplate reader capable of measuring absorbance at 450 nm
- Pipettes and pipette tips
- Deionized or distilled water
- Polypropylene tubes for dilution of standard
- Paper towels or laboratory wipes
- 1N H_2SO_4 or 1N HCl
- Bovine Serum Albumin Fraction V (BSA)
- Tris(hydroxymethyl)aminomethane (Tris)
- Sodium Chloride (NaCl)

8. Technical Hints

- Samples generating values higher than the highest standard should be further diluted in the appropriate sample dilution buffers.
- Avoid foaming or bubbles when mixing or reconstituting components.
- Avoid cross contamination of samples or reagents by changing tips between sample, standard and reagent additions.
- Ensure plates are properly sealed or covered during incubation steps.
- Complete removal of all solutions and buffers during wash steps is necessary to minimize background.
- All samples should be mixed thoroughly and gently.
- Avoid multiple freeze/thaw of samples.
- Incubate ELISA plates on a plate shaker during all incubation steps.

9. Reagent Preparation

- Equilibrate all reagents to room temperature (18-25°C) prior to use.
- Prepare only as much reagent as is needed on the day of the experiment.

9.1 TBS buffer:

Prepare 0.1M Tris, 0.15M NaCl, pH 7.4.

9.2 Blocking buffer (BB):

Prepare 3% BSA (w/v) in TBS.

9.3 1X Wash Buffer:

Dilute 50 mL of 10X wash buffer concentrate with 450 mL of deionized water.

10. Sample Preparation

- Use rat plasma (citrate, heparin, EDTA) samples with this assay.
- Centrifuge for 15 mins at 1000 *xg* within 30 mins of collection.
- Assay immediately or aliquot and store at ≤-20°C. Avoid repeated freezing and thawing.

11. Sample Dilution

- The assay measures Prolactin antigen in the 2-1000 ng/ml range.
- Samples with rat prolactin levels above 1000ng/mL should be diluted in blocking buffer before use.
- Normal plasma should not require dilution before use in this assay.

12. Preparation of Standards

– Dilutions for the standard curve and zero standard must be made and applied to the plate immediately.

12.1 Reconstitute standard by adding 1 ml of blocking buffer directly to the vial and agitate gently to completely dissolve contents. This will result in a 1000 ng/ml plasma standard.

12.2 Then dilute using Blocking buffer (BB) according to the table below:

Prolactin concentration (ng/ml)	Dilutions
1000	From vial
500	500µl (BB) + 500µl (1000 ng/ml)
200	600µl (BB) + 400µl (500 ng/ml)
100	500µl (BB) + 500µl (200 ng/ml)
50	500µl (BB) + 500µl (100 ng/ml)
20	600µl (BB) + 400µl (50 ng/ml)
10	500µl (BB) + 500µl (20 ng/ml)
5	500µl (BB) + 500µl (10 ng/ml)
2	600µl (BB) + 400µl (5 ng/ml)
0	500µl (BB) Zero point to determine background

13. Assay Procedure

- Equilibrate all materials and prepared reagents to room temperature prior to use.
- Vigorously shake plate (300 rpm) at each step of the assay.

13.1 Standard and Unknown Addition.

13.1.1 Remove microtiter plate from bag and add 100 µL Prolactin standards (in duplicate) and unknowns to wells.

13.1.2 Carefully record position of standards and unknowns. Shake plate at 300 rpm for 30 mins.

13.1.3 Wash wells three times with 300 µL wash buffer. Remove excess wash by gently tapping plate on paper towel.
Δ Note: The assay measures total rat prolactin in the 2-1000 ng/mL range. Samples with rat prolactin levels above 1000ng/mL should be diluted in blocking buffer before use. Normal plasma should not require dilution before use in this assay.

13.2 Primary Antibody Addition.

13.2.1 Reconstitute primary antibody by adding 10 mL of blocking buffer directly to the vial and agitate gently to completely dissolve contents.

13.2.2 Add 100 µL to all wells. Shake plate at 300 rpm for 30 mins.

13.2.3 Wash wells three times with 300 µL wash buffer. Remove excess wash by gently tapping plate on paper towel.

13.3 Anti-Rabbit HRP Secondary Reagent Addition.

13.3.1 Briefly centrifuge vial before opening.

13.3.2 Dilute 2 µL of anti-rabbit HRP secondary reagent into 10 mL blocking buffer to generate a 1:5,000 dilution.

13.3.3 Add 100 µL of the 1:5,000 dilution to all wells. Shake plate at 300 rpm for 30 mins.

13.3.4 Wash wells three times with 300 µL wash buffer. Remove excess wash by gently tapping plate on paper towel.

13.4 Substrate Incubation.

13.4.1 Add 100 µL TMB substrate to all wells and shake plate for 2 to 10 mins. Substrate will change from colorless to different strengths of blue.

13.4.2 Quench reaction by adding 50 µL of 1N H_2SO_4 or HCl stop solution to all wells when samples are visually in the same

range as the standards. Add stop solution to wells in the same order as substrate upon which color will change from blue to yellow. Mix thoroughly by gently shaking the plate.

13.5 Measurement.

13.5.1 Set the absorbance at 450 nm in a microtiter plate spectrophotometer.

13.5.2 Measure the absorbance in all wells at 450 nm.

13.5.3 Subtract zero point from all standards and unknowns to determine corrected absorbance (A_{450}).

13.6 Calculation of Results.

13.6.1 Plot A_{450} against the amount of prolactin in the standards.

13.6.2 Fit a straight line through the linear points of the standard curve using a linear fit procedure if unknowns appear on the linear portion of the standard curve.

13.6.3 Alternatively, create a standard curve by analyzing the data using a software program capable of generating a four parameter logistic (4PL) curve fit.

13.6.4 The amount of Prolactin in the unknowns can be determined from this curve.

13.6.5 If samples have been diluted, the calculated concentration must be multiplied by the dilution factor.

14. Typical Results

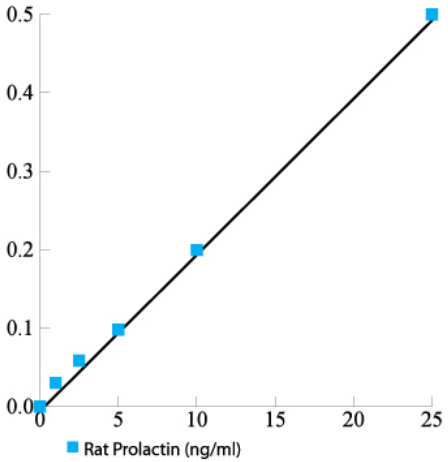

Fig 1. A typical standard curve. Example only.

15. Performance Characteristics

Expected Values:

The concentration of prolactin in pooled normal rat plasma determined in house testing was 0.8-3.0 ng/mL. The concentration in control rat plasma as determined by radioimmunoassay was 10.5 ng/ml Prolactin levels in pregnant rats are elevated immediately before birth.

Sensitivity:

The minimum detectable dose (MDD) was determined by adding two standard deviations to the mean optical density value of eighteen zero standard replicates (range OD450: 0.066-0.078) and calculating the corresponding concentration.

The MDD was 0.168 µg/ml.

16. Specificity

Pooled normal plasma from pig, dog, human and sheep was assayed and no cross-reactivity was observed. Pooled normal mouse and horse plasma showed significant cross-reactivity. Pooled normal plasma from rhesus monkey, cyno monkey and rabbit resulted in significant background color development.

17. Notes

Technical Support

For all technical or commercial enquiries please go to:

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