

ab282916 – Trypsin Activity Assay Kit (Fluorometric)

For the measurement of trypsin activity in Biological Fluids, Tissues.

For research use only - not intended for diagnostic use.

For overview, typical data and additional information please visit:

<http://www.abcam.com/ab282916>

Storage and Stability

Store entire kit at -20°C, protected from light. Upon opening, use within 2 months.

Materials Supplied

Item	Quantity	Storage Condition
Trypsin Assay Buffer	35 ml	-20°C
Trypsin Substrate	50 µl	-20°C
Positive control (lyophilized)	1 vial	-20°C
AMC standard	100 µl	-20°C
Trypsin Inhibitor	500 µl	-20°C

Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully utilize this assay:

- 96-well clear plate with flat bottom
- Dounce Tissue Homogenizer
- Multi-well spectrophotometer

Reagent Preparation Before using the kit, spin the tubes prior to opening.

Trypsin Assay Buffer: Store at either 4°C or -20°C. Bring to room temperature (RT) before use.

Trypsin Substrate: Store at -20°C. Protect from light. Bring to RT before use.

Positive Control (lyophilized): Reconstitute with 100 µl Trypsin Assay Buffer. Pipette up and down to dissolve completely. Aliquot and store at -20°C. Keep on ice while in use.

AMC Standard (1 mM in DMSO): Store at -20°C. Bring to RT before use.

Trypsin Inhibitor (TLCK, 20 mM): Store at -20°C. Bring to RT before use.

Sample Preparation

Serum Sample(s):

1. Clarify Serum Sample by centrifugation at 1,000 x g and 4°C for 10 min in order to reduce the turbidity and separate the insoluble materials.
2. Prepare a 20-fold dilution of the Serum Sample in Trypsin Assay Buffer (10 µl of Serum with 190 µl of Trypsin Assay Buffer) and incubate at 60°C for 20 min.
3. Centrifuge at 10,000 x g and 4°C for 10 min and transfer the supernatant to a fresh tube.
4. Add 40-70 µl of the diluted Sample into two parallel wells of a 96-well clear plate labeled as "Sample" and "Sample Background Control".
5. Add 10 µl of DMSO into the Sample well and 10 µl of Trypsin Inhibitor into the Sample Background Control well respectively.
6. Adjust the volume of the Sample and Sample Background Control well to 80 µl/well with Trypsin Assay Buffer. Incubate at RT for 10 min in dark.

Tissue Sample(s):

1. Add 250 µl of ice-cold Trypsin Assay Buffer to 50 mg of the Tissue Sample (wet weight).
2. Homogenize on ice using a Dounce Tissue homogenizer.
3. Centrifuge the Sample(s) at 12,000 x g and 4°C for 10 min. Collect the supernatant.
4. Incubate the supernatant at 60°C for 20 min. Centrifuge at 10,000 x g at 4°C for 10 min and collect the Sample supernatant. Add 2-20 µl of the Sample supernatant into well(s) of a 96-well clear plate labeled as "Sample".
5. Prepare an additional well with the same volume of Trypsin Assay Buffer labeled as "Background Control" by adding 80 µl of Trypsin Assay Buffer.
6. Adjust the volume of the Sample well(s) to 80 µl/well with Trypsin Assay Buffer.

Positive Control:

1. Prepare an 800-fold dilution of the Positive Control (1 µl of Positive Control and 799 µl of Trypsin Assay Buffer).
2. Add 2-6 µl of the diluted Positive Control into desired wells(s). Adjust the volume of the Positive Control well to 80 µl/well with Trypsin Assay Buffer.

Note: For Unknown Samples, we recommend doing a pilot experiment and testing several doses to ensure that the readings are within the linear range of the Standard Curve.

Standard Curve Preparation

1. Prepare 0.1 mM AMC Standard solution by diluting the 1 mM stock AMC Standard (10 µl of 1 mM stock AMC Standard and 90 µl of Trypsin Assay Buffer).
2. Add 0, 2, 4, 6, 8, 10 µl of 0.1 mM (0.1 nmol/µl) AMC Standard solution into a series of wells of a 96-well clear plate to generate 0, 0.2, 0.4, 0.6, 0.8, 1 nmol/well of AMC Standard respectively.
3. Adjust the volume of each well to 100 µl with Trypsin Assay Buffer.

Substrate Mix Preparation

1. Prepare a 40-fold dilution of the Trypsin Substrate stock solution (2 µl of Trypsin Substrate and 78 µl Trypsin Assay Buffer resp.), vortex briefly.
2. Add 20 µl of the diluted Trypsin Substrate solution to each well containing Sample(s), Positive Control and Background Control wells. The total volume in every well including Standards, Sample(s), Positive Control & Background Controls should be 100 µl.

Measurement

1. Measure the fluorescence intensity (Ex/Em = 380/460 nm) in kinetic mode for 10-60 min at RT using a fluorescence microtiter plate reader.
2. Choose any two time points (t_1 & t_2) in the linear range of the plot and obtain the corresponding RFU for all Samples (RS_1 and RS_2) and Background Controls (RB_1 and RB_2).
3. The AMC Standard Curve can be read in endpoint mode.

Note: Shake the microplate carefully for 5 sec to mix the contents prior to reading the plate.

Calculation

1. Subtract 0 Standard RFU reading from all Standard readings.
2. Plot the AMC Standard Curve and obtain the slope of the curve (RFU/nmol).
3. Apply Samples Δ RFU (RS_2-RS_1) and Sample Background Controls Δ RFU (RB_2-RB_1) for serum samples or Samples Δ RFU (RS_2-RS_1) and Background Control Δ RFU (RB_2-RB_1) for tissue samples to the AMC Standard Curve to obtain the corresponding amount of AMC formed during the reaction time ($\Delta t = t_2-t_1$).
4. Calculate the Background-corrected Sample (B, in nmol) by subtracting the amount of AMC formed by Sample Background Control and/or Background Control from the amount of AMC formed by Sample.
5. Calculate Trypsin Activity in the Sample as:

$$\text{Sample Trypsin Activity} = \frac{\text{B Sample(corrected)}}{1\Delta t \cdot V \cdot P} * D = \text{nmol/min/mg} = \text{mU/mg}$$

Where:

B is background-corrected AMC amount from the AMC Standard Curve (nmol)

Δt is the Reaction time (t_2-t_1 in min)

V is the Sample volume added into the reaction well (ml)

P is the initial protein concentration (mg/ml)

D is the Dilution factor (D = 1 for undiluted Samples)

Unit Definition: One unit of Trypsin activity is the amount of enzyme that generates 1.0 μ mol of AMC per min at pH 8.0 at RT.

Technical Support

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