

ab283361 - Ornithine Decarboxylase/ODC Inhibitor Screening Kit (Colorimetric)

For screening for inhibitors of human Ornithine decarboxylase I (ODC1)
For research use only - not intended for diagnostic use.

For overview, typical data and additional information please visit:

<http://www.abcam.com/ab283361>

Storage and Stability

On receipt entire assay kit should be stored at -20°C, protected from light. Upon opening, use kit within 1 year.

Materials Supplied

Item	Quantity	Storage Condition
ODC1 Assay Buffer	25 mL	-20°C
ODC1 Substrate	1 vial	-20°C
ODC1 Converter Mix	1 vial	-20°C
ODC1 Enzyme Mix	1 vial	-20°C
ODC1 Cofactor	1 vial	-20°C
Human ODC1	1 vial	-20°C
DFMO (in DMSO)	20 µl	-20°C

Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully utilize this assay:

- 96-well UV-transparent plate with flat bottom
- Multi-well spectrophotometer (ELISA reader)
- 1 M DTT
- 50%

Glycerol

Reagent Preparation

- Before using the kit, spin the tubes prior to opening.

ODC1 Assay Buffer: Warm to room temperature before use. Store at 4°C or -20°C.

ODC1 Substrate, ODC1 Converter: Reconstitute each vial with 220 µl dH₂O. Pipette up and down to dissolve. Store at -20°C.

ODC1 Enzyme Mix: Reconstitute with 220 µl ODC1 Assay Buffer. Pipette up and down to dissolve. Aliquot and store at -20°C. Avoid repeated freeze/thaw cycles. Keep on ice while in use.

ODC1 Cofactor: Reconstitute with 1.2 ml dH₂O. Pipette up and down to dissolve. Store at -20°C. Avoid light. Keep on ice while in use.

Human ODC1: *Only reconstitute prior to use!* Mix 5 µl of 1 M DTT solution with 995 µl of 50% glycerol solution (not provided). Add 40 µl of the DTT/50% glycerol solution into the vial. Vortex for 5 seconds. Incubate at 25 °C for 30 minutes. Completely dissolved Human ODC1 should be a viscous clear yellow solution. Aliquot and store at -80°C. Avoid freeze and thaw. Use within two months.

DFMO (in DMSO): Ready to use as supplied. Store at -20°C. Warm to room temperature before use.

Screening Protocol

Test compounds, Inhibitor Control, Enzyme Control & Background Control Preparations:

- Dissolve candidate inhibitors at 100X highest final test concentration into an appropriate solvent.
- Dilute to 10X the desired test concentration with ODC1 Assay Buffer.
- Sample compound [S]:** Add 10 µl diluted test inhibitor into a 96-well UV-transparent plate (not provided).
- Enzyme Control [EC]:** Add 10 µl of Assay Buffer to a designated well(s).
- Inhibitor control [IC]:** Dilute DFMO by adding 2 µl of the stock solution into 198 µl of ODC1 Assay Buffer.
- Add 10 µl of the diluted DFMO inhibitor into designated well(s).
- Background Control [BC]:** Add 10 µl of the diluted DFMO in a well designated as Background Control [BC].

ΔNote: If the inhibitor sample or solvent has significant absorbance at 340 nm, add 10 µl diluted test inhibitor in a parallel well designated as Sample Control [SC]. Up to 10% DMSO does not affect the reaction. However, it is recommended to study solvent effects by adding the same amount of solvent into parallel well(s) designated as Solvent Control [SolC].

Enzyme Solution Preparation:

- Mix enough reagents for the number of assays to be performed.
- Prepare a 100-fold dilution of the Human ODC1 (e.g. Mix 2 µl of Human ODC1 with 198 µl ODC1 Assay Buffer. 50% glycerol solution is viscous. Handle human ODC1 solution carefully.
- For each well, prepare 40 µl ODC1 Enzyme Solution:

Item	Amount
ODC1 Assay Buffer	20 µl
Diluted Human ODC1	20 µl

- Mix and add 40 µl of the ODC1 enzyme solution to Sample, Inhibitor Control, Enzyme Control and Solvent Control wells ([S], [IC], [EC] and [SolC]).
- Add 40 µl of Assay Buffer into Background Control and Sample control ([BC] and [SC]) well(s). Mix well, and incubate the plate for 5 min at 37 °C.

ΔNote: Do not store unused diluted DFMO solutions.

Reaction Mix:

Mix enough reagents for the number of assays to be performed. For each well, prepare 50 µl Mix containing:

Item	Reaction Mix
ODC1 Assay Buffer	34 µl
ODC1 Substrate	2 µl
ODC1 Converter	2 µl
ODC1 Enzyme Mix	2 µl
ODC1 Cofactor	10 µl

Mix and add 50 µl of the Reaction Mix to all wells ([S], [EC], [IC], [SC], [BC], [SolC]). Mix well.

Measurement:

Immediately, measure absorbance (OD: 340 nm) in a microplate reader in kinetic mode at 37°C for 20-30 min, taking measurement readings every 20 seconds.

Calculation:

1. Take the linear portion of the kinetic curve $\Delta OD = OD_{\text{end}} - OD_{\text{initial}}$ and divide by reaction time ($\Delta t = t_{\text{end}} - t_{\text{initial}}$) to get the rate of individual well (note ΔOD rates would be negative).
2. Subtract the Background Control [BC] rate from all readings to obtain activity for each reading (if Sample Control [SC] is higher than Background Control [BC] for a given test compound, subtract its rate from the signal of that particular sample only).
3. Set the activity of Enzyme Control [EC] as 100% (in case Solvent Control is significantly different from EC, replace with [SolC] values in the formulas below).
4. Calculate % Inhibition or % Relative Activity of the test inhibitors as follows:

$$\% \text{ Inhibition} = \frac{\text{Activity of EC} - \text{Activity of S}}{\text{Activity of EC}} \times 100$$

$$\% \text{ Relative activity} = \frac{\text{Activity of S}}{\text{Activity of EC}} \times 100$$

ΔNote: A slow kinetic response in the reactions (Lag phase) may be observed. Do not use the first 3-5 minutes of the reaction curves for the estimation of ODC activity.

Technical Support

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