ab284021– Human S100A16 ELISA Kit

For the quantitative measurement of S100A16 in human cell lysate samples.
For research use only - not intended for diagnostic use.

For overview, typical data and additional information please visit:
http://www.abcam.com/ab284021

Storage and Stability: Store kit at +4°C immediately upon receipt, apart from the SP Conjugate and Biotinylated Antibody which should be stored at -20°C. Kit has a storage time of 1 year from receipt, providing components have not been reconstituted.

Materials Supplied

Item	Quantity	Storage Condition
Human S100A16 Microplate	96 wells	4°C
Sealing Tapes	3	N/A
100X Streptavidin-Peroxidase Conjugate	80 µL	-20°C
10X Diluent N Concentrate	30 mL	4°C
1X Standard Diluent	2 mL	4°C
20X Wash Buffer Concentrate	2 x 30 mL	4°C
50X Biotinylated Human S100A16 Antibody	1 vial	-20°C
Chromogen Substrate	7 mL	4°C
S100A16 Standard	1 vial	-20°C
Stop Solution	11 mL	4°C

Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully utilize this assay:

Microplate reader capable of measuring absorbance at 450 nm
Pipettes (1-20 µL, 20-200 µL, 200-1000 µL, and multiple channel)
Deionized or distilled reagent grade water

Reagents Preparation

Equilibrate all reagents to room temperature (18-25°C) prior to use. The kit contains enough reagents for 96 wells.
Prepare only as much reagent as is needed on the day of the experiment.
If crystals have formed in the concentrate, mix gently until the crystals have completely dissolved.
Δ Note: Concentration of the kit components are lot-specific, and the end user should always refer to the vial label.

10X Diluent N Concentrate: Dilute the Diluent N Concentrate 10-fold with reagent grade water to produce a 1x solution. Store for up to 30 days at 2-8°C.

50X Biotinylated Human S100A16 Antibody: Spin down the antibody briefly and dilute the desired amount of the antibody 50-fold with Diluent N to produce a 1x solution. The undiluted antibody should be stored at -20°C.

20X Wash Buffer Concentrate: Dilute the Wash Buffer Concentrate 20-fold with reagent grade water to produce a 1x solution.

100X Streptavidin-Peroxidase Conjugate: Spin down the SP Conjugate briefly and dilute the desired amount of the conjugate 100-fold with Diluent N to produce a 1x solution. The undiluted conjugate should be stored at -20°C.

Standard Preparation

Always prepare a fresh set of standards for every use.
Prepare serially diluted standards immediately prior to use.
Any remaining standard should be stored at -20°C after reconstitution and used within 30 days.
The following section describes the preparation of a standard curve for duplicate measurements (recommended).
Reconstitution of the Human S100A16 Standard vial to prepare 40 ng/mL Stock Standard.

- First consult the Human S100A16 Standard vial to determine the mass of protein in the vial.
- Calculate the appropriate volume of 1X Diluent N to add when resuspending the Human S100A16 Standard vial to produce 40 ng/mL Stock Standard by using the following equation:
  - C_S = Starting mass of Human S100A16 Standard (see vial label) (µg)
  - C_F = The 40 ng/mL S100A16 Stock Standard final required concentration
  - V_D = Required volume of 1X Diluent N for reconstitution (µL)
  - Calculate total required volume 1X Diluent N for resuspension:

$$(C_S / C_F) \times 1,000 = V_D$$

Example: **Δ Note: This example is for demonstration purposes only. Please remember to check your standard vial for the actual amount of standard provided**.

CS = 125 ng of Human S100A16 Standard in vial

CF = 40 ng/mL Human S100A16 Standard #1 final concentration

VD = Required volume of 1X Diluent N for reconstitution (125 ng / 40 ng/mL) x 1,000 = 3125 µL

- Reconstitute the Human S100A16 Standard vial by adding the appropriate calculated amount VD of 1X Diluent N to the vial to generate the 40 ng/mL Human S100A16 Standard #1. Mix gently and thoroughly.
- Allow the reconstituted 125 ng/mL Albumin Standard #1 to sit for 10 minutes with gentle agitation prior to making subsequent dilutions
- Label five tubes #2 – 8.
- Add 120 µL of 1X Diluent N to tube #2 – 8.
- To prepare Standard #2, add 120 µL of the Standard #1 into tube #2 and mix gently.
- To prepare Standard #3, add 120 µL of the Standard #2 into tube #3 and mix gently
- Using the table below as a guide, prepare subsequent serial dilutions. 1X Diluent N serves as the zero standard (0 ng/mL).

Standard #	Volume to dilute (µL)	Volume 1X Diluent N (µL)	S100A16 (ng/mL)
1	240 µL	-	40
2	120 µL Standard #1	120	20
3	120 µL Standard #2	120	10
4	120 µL Standard #3	120	5
5	120 µL Standard #4	120	2.5
6	120 µL Standard #5	120	1.25
7	120 µL Standard #6	120	0.625
8	-	120	0.0

Sample Preparation

Cell Lysate: Rinse cell with cold PBS and then scrape the cell into a tube with 5 mL of cold PBS and 0.5 M EDTA. Centrifuge suspension at 1500 rpm for 10 minutes at 4°C and aspirate supernatant. Re-suspend pellet in ice-cold Lysis Buffer (10 mM Tris, pH8.0, 130 mM NaCl, 1% Triton X-100, protease inhibitor cocktail). For every 1 x 106 cells, add approximately 100 µl of ice-cold Lysis Buffer. Incubate on ice for 60 minutes. Centrifuge at 13000 rpm for 30 minutes at 4°C and collect supernatant.

Applicable samples may also include biofluids, cell culture, and tissue homogenates. If necessary, user should determine optimal dilution factor depending on application needs.

Plate Preparation

The 96 well plate strips included with this kit are supplied ready to use. It is not necessary to rinse the plate prior to adding reagents.
Unused plate strips should be immediately returned to the foil pouch containing the desiccant pack, resealed, and stored at 4°C.
For each assay performed, a minimum of two wells must be used as the zero control.
For statistical reasons, we recommend each sample should be assayed with a minimum of two replicates (duplicates).

Assay Procedure

Equilibrate all materials and prepared reagents to room temperature prior to use.
We recommend that you assay all standards, controls and samples in duplicate. The assay is performed at room temperature (20-25°C).

1. Add 50 µL of Human S100A16 Standard or sample to each well. Gently tap plate to thoroughly coat the wells. Break any bubbles that may have formed. Cover wells with a sealing tape and incubate for 2 hours. Start the timer after the last addition.
2. Wash five times with 200 µL of Wash Buffer manually. Invert the plate each time and decant the contents; hit 4-5 times on absorbent material to completely remove the liquid. If using a machine, wash six times with 300 µL of Wash Buffer and then invert the plate, decanting the contents; hit 4-5 times on absorbent material to completely remove the liquid.
3. Add 50 µL of Biotinylated Human S100A16 Antibody to each well. Gently tap plate to thoroughly coat the wells. Break any bubbles that may have formed. Cover wells with a sealing tape and incubate for 1 hour.
4. Wash the microplate as described above.
5. Add 50 µL of SP Conjugate to each well. Gently tap plate to thoroughly coat the wells. Break any bubbles that may have formed. Cover wells with a sealing tape and incubate for 30 minutes. Turn on the microplate reader and set up the program in advance.
6. Wash the microplate as described above.
7. Add 50 µL of Chromogen Substrate to each well. Gently tap plate to thoroughly coat the wells. Break any bubbles that may have formed. Incubate for 30 minutes or until the optimal blue colour density develops.
8. Add 50 µL of Stop Solution to each well. The colour will change from blue to yellow. Gently tap plate to ensure thorough mixing. Break any bubbles that may have formed.
9. Read the absorbance on a microplate reader at a wavelength of 450 nm immediately. If wavelength correction is available, subtract readings at 570 nm from those at 450 nm to correct optical imperfections. Otherwise, read the plate at 450 nm only. Please note that some unstable black particles may be generated at high concentration points after stopping the reaction for about 10 minutes, which will reduce the readings.

Download our ELISA guide for technical hints, results, calculation, and troubleshooting tips:
www.abcam.com/protocols/the-complete-elisa-guide
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Additional information

CALCULATION

1. Calculate the mean value of the duplicate or triplicate readings for each standard and sample.
2. To generate a standard curve, plot the graph using the standard concentrations on the x-axis and the corresponding mean 450 nm absorbance (OD) on the y-axis. The best fit line can be determined by regression analysis using log-log or four-parameter logistic curve fit.
3. Determine the unknown sample concentration from the Standard Curve and multiply the value by the dilution factor.

TYPICAL DATA

Typical data provided for demonstration purposes only. A new standard curve must be generated for each assay performed.

Standard Point	ng/mL	OD	Average OD
P1	40	1.857 1.795	1.826
P2	20	1.529 1.520	1.525
P3	10	1.017 0.997	1.007
P4	5.0	0.585 0.575	0.580
P5	2.5	0.334 0.329	0.332
P6	1.25	0.230 0.200	0.202
P7	0.625	0.160 0.106	0.133
P8	0.0	0.070 0.066	0.068

PERFORMANCE CHARACTERISTICS

1. The minimum detectable dose of human S100A16 as calculated by 2SD from the mean of a zero standard was established to be 0.4 ng/mL.

RECOVERY

Standard Added Value	2.5 - 20 ng/mL
Recovery %	87 – 114
Average Recovery %	96

CROSS REACTIVITY

Protein	Cross-Reactivity (%)
S100A3	20

Download our ELISA guide for technical hints, results, calculation, and troubleshooting tips:
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