

ab286867 – Cibacron Blue-Agarose Beads

For purification of a variety of Cibacron Blue-binding proteins.

For research use only - not intended for diagnostic use.

For overview, typical data and additional information please visit:

<http://www.abcam.com/ab286867>

Storage and Stability

Store at 4°C. Do not freeze the resin.

Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully utilize this assay:

- Binding buffer e.g. 50 mM Tris buffer, pH 8.0 or PBS.
- Elution buffer: e.g. 50 mM Tris, 1 M NaCl, pH 8.0 or another buffer containing a competing ligand.

Column Preparation

Carefully pack 1-5 ml of resin slurry in a disposable column avoiding air bubbles and allow the buffer to drain through. Wash the resin with 4x 5 column volumes (CV) of Binding Buffer. Do not allow the resin to dry. Close the column outlet.

Purification Protocol

1. Load sample on the resin (If necessary dilute to 5 ml with binding buffer). Close the column with a cap.
2. Allow the sample to bind to the resin for 30 min by mixing the suspension on a rotary shaker or intermittently by hand.
3. After 30 min, open the column (both top and outlet) and collect the flow-through fraction.
4. Wash the column with 5-10 CV of binding buffer. Combine washes together and concentrate if necessary.
5. Elute the protein from the resin using appropriate elution buffer.
6. Analyze the flow through, wash and eluted protein, by SDS-PAGE, UV or any other functional assay.
7. To regenerate/store column: a) Wash with 5 volumes of Elution Buffer; b) Wash with 5 volumes of distilled water; c) Store column in 20 % Ethanol/H₂O at 4°C.

To clean/store column

1. Wash with 5 volumes of 0.2 N NaOH or 6 M Guanidine HCl.
2. Wash with 5 volumes of distilled water.
3. Store column in 20 % Ethanol/H₂O at 4°C.

Technical Support

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