

ab291062 - Hamster (CHO) GLUTATHIONE S-TRANSFERASE PI (GSTp)

ELISA Kit

For Determination of GSTp in Hamster (CHO) Samples.
For research use only and is not intended for diagnostic use.

For overview, typical data and additional information please visit:

<http://www.abcam.com/ab291062>

Storage and Stability

The expiration date for the kit and its components is stated on the box label. All components should be stable up to the expiration date if stored and used per this kit protocol insert. On receipt entire assay kit should be stored at 4-8°C, protected from light.

Materials Supplied

Item	Quantity	Storage Condition
ELISA Micro Plate, antibody coated	12 X 8 unit	4-8°C
Detection Antibody 100X	150 µL	4-8°C, store in dark
HRP-Streptavidin 100X	150 µL	4-8°C, store in dark
Calibrator	1 vial	4-8°C
Diluent Solution	60 mL	4-8°C
20 X Wash Solution Concentrate	50 mL	4-8°C
Chromogen Substrate Solution	12 mL	4-8°C, store in dark
STOP Solution	12 mL	4-8°C

Materials Required, Not Supplied

These material are not included in the kit but will be required to successfully utilize this assay:

- Precision pipettes (2 µL to 100 µL) for making and dispensing dilutions
- Test tubes
- Squirt bottle or Microtitre washer/aspirator
- Distilled or Deionized H₂O
- Microtitre Plate reader
- Assorted glassware for the preparation of reagents and buffer solutions
- Centrifuge for sample collection
- Anticoagulant for plasma collection
- Timer
- Microplate shaker

Reagent Preparation

- Bring all reagents to room temperature (16°C to 25°C) before use.

Diluent Solution – Ready to use as supplied.

Wash Solution Concentrate - Dilute 1/20 with distilled or deionized water (1 part buffer concentrate, 19 parts dH₂O). Crystal formation in the concentrate may occur when storage temperatures are low. Warming of the concentrate to 30-35°C before dilution can dissolve crystals.

Detection Antibody - Calculate the required amount of working conjugate solution for each microtitre plate test strip by adding 10 µL Detection Antibody to 990 µL of 1X Diluent Solution for each test strip to be used for testing. Dilute immediately before use and protect from light. Mix uniformly, but gently. Avoid foaming.

HRP-Streptavidin - Calculate the required amount of working conjugate solution for each microtitre plate test strip by adding 10 µL HRP-Streptavidin to 990 µL of 1X Diluent Solution for each test strip to be used for testing. Dilute immediately before use and protect from light. Mix uniformly, but gently. Avoid foaming.

Pre-coated ELISA Micro Plate - Ready to use as supplied. Unseal foil pouch and remove plate from pouch. Remove all strips and wells that will not be used in the assay and place back in pouch and re-seal along with desiccant.

Hamster (CHO) GSTp Calibrator – **NOTE: The calibrator is provided at the concentration stated on the vial.** Reconstitute the Calibrator with distilled or de-ionized water as specified on the vial and mix gently until dissolved. The amount of calibrator is shown on the vial, and after reconstitution will have a concentration of X µg/mL, where X is the amount on the vial (the reconstituted calibrator should be aliquoted and stored frozen if future use is intended).

All biological materials should be handled as potentially hazardous. Follow universal precautions when handling and disposing.

Known interfering substances - Azide and thimerosal at concentrations higher than 0.1% inhibits the enzyme reaction.

Standard preparation

Prepare the Following Standards:

Standard 0 (0.0 ng/mL) in duplicate

Standard 1 (0.63 ng/mL) in duplicate

Standard 2 (1.25 ng/mL) in duplicate

Standard 3 (2.50 ng/mL) in duplicate

Standard 4 (5 ng/mL) in duplicate

Standard 5 (10 ng/mL) in duplicate

Standard 6 (20 ng/mL) in duplicate

NOTE: Please remember to check your calibrator vial for the actual concentration of calibrator provided.

Sample preparation

Sample Collection and Handling

The assay requires that each test sample be diluted before use. All samples should be assayed in duplicate each time the assay is performed. The recommended dilutions are

only suggestions. Dilutions should be based on the expected concentration of the unknown sample such that the diluted sample falls within the dynamic range of the standard curve. If unsure of sample level, a serial dilution with one or two representative samples before running the entire plate is highly recommended. Dilute samples immediately prior use.

- **CHO culture extract samples** – Recommended starting dilution is 1/20. To prepare a 1/20 dilution of a sample, transfer 15 µL of sample to 285 µL of 1X diluent. This gives you a 1/20 dilution. Mix thoroughly.

Assay Protocol

- Bring all reagents and samples to room temperature 30 minutes prior to the assay.
 - All standards and samples should be run in duplicate.
1. The Standards and the test sample(s) should be loaded into the ELISA wells as quickly as possible to avoid a shift in OD readings. Using a multichannel pipette would reduce this occurrence.
 2. Pipette 100 µL of:

Standard 0	(0.0 ng/mL) in duplicate
Standard 1	(0.63 ng/mL) in duplicate
Standard 2	(1.25 ng/mL) in duplicate
Standard 3	(2.50 ng/mL) in duplicate
Standard 4	(5.0 ng/mL) in duplicate
Standard 5	(10.0 ng/mL) in duplicate
Standard 6	(20.0 ng/mL) in duplicate

3. Pipette 100 µL of sample (in duplicate) into pre-designated wells.
4. Incubate the micro titer plate while shaking on a microplate shaker at 400 rpm at room temperature for a hundred and twenty (120 ± 2) minutes. Keep plate covered and level during incubation.
5. Following incubation, aspirate the contents of the well.
6. Completely fill each well with appropriately diluted Wash Solution and aspirate. Repeat three times, for a total of four washes. If washing manually: completely fill wells with wash buffer, invert the plate then pour/shake out the contents in a waste container. Follow this by sharply striking the wells on absorbent paper to remove residual buffer. Repeat 3 times for a total of four washes.
7. Pipette 100 µL of appropriately diluted Enzyme-Antibody Conjugate to each well. Incubate at room temperature for thirty (30 ± 2) minutes. Keep plate covered in the dark and level during incubation.
8. Wash and blot the wells as described in Steps 5/6.
9. Pipette 100 µL of TMB Substrate Solution into each well.
10. Incubate in the dark at room temperature for precisely ten (10) minutes.
11. After ten minutes, add 100 µL of Stop Solution to each well.
12. Determine the absorbance (450 nm) of the contents of each well within 30 minutes. Calibrate the plate reader to manufacturer's specifications.

Calculation

- Subtract the average background value (Average absorbance reading of Standard zero) from the test values for each sample.
- Average the duplicate readings for each standard and use the results to construct a Standard Curve. Construct the standard curve by reducing the data using computer software capable of generating a four parameter logistic curve fit. A second order polynomial (quadratic) or other curve fits may also be used; however, they will be a less precise fit of the data.
- Interpolate test sample values from standard curve. Correct for sera dilution factor to arrive at the GSTp concentration in original samples.

Technical Support

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