

## ab324118 – Enterokinase Activity Assay Kit (Colorimetric)

A sensitive assay for quantifying enterokinase activity.  
For research use only - not intended for diagnostic use.

For overview, typical data and additional information please visit: [www.abcam.com/ab324118](http://www.abcam.com/ab324118)

**Storage and Stability:** Store kit at -20°C in the dark immediately upon receipt. Refer to list of materials supplied for storage conditions of individual components.

### Materials Supplied

Item	Quantity	Storage Condition
EK Yellow™	1 vial	-20°C (Store in the dark)
Enterokinase Substrate	1 vial	-20°C (Store in the dark)
Assay Buffer	1 bottle (10 mL)	-20°C
Enterokinase Standard	1 vial	-20°C (Store in the dark)
DMSO	1 vial (100 µL)	-20°C

### Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully utilize this assay:

- Absorbance microplate reader capable of measuring at 405 nm (Path check on)
- Clear bottom plates
- ddH<sub>2</sub>O
- 0.1% BSA

### Protocol Summary

1. Prepare test samples and enterokinase standards (50 µL).
2. Add equal volume of Enterokinase working solution (50 µL).
3. Incubate at 37 °C for 30 - 60 minutes.
4. Monitor absorbance increase at 405 nm.

**IMPORTANT:** Thaw one vial of each kit component at room temperature before starting the experiment.

### Preparation of Stock Solutions

Unless otherwise noted, all unused stock solutions should be divided into single-use aliquots and stored at -20 °C after preparation. Avoid repeated freeze-thaw cycles.

#### EK Yellow™ stock solution (100X)

1. Add 50 µL of DMSO into EK Yellow™ to make 100X stock solution.

#### Enterokinase Substrate stock solution (100X)

1. Add 50 µL of DMSO into Enterokinase Substrate to make 100X stock solution.

#### Enterokinase standard solution (10 µg/mL)

1. Add 50 µL of ddH<sub>2</sub>O + 0.1% BSA into Enterokinase Standard vial to make 10 µg/mL Enterokinase stock solution.

### Preparation of Standard Solution

#### Enterokinase standard

1. Add 10 µL of 10 µg/mL Enterokinase standard solution into 990 µL of Assay Buffer to get 100 ng/mL enterokinase solution (EK7). Then perform 1:2 serial dilutions in assay buffer to get serially diluted enterokinase standards (EK6 - EK1).

**Note:** The EK standards are for positive control only, and should not be relied on as a quantitation standard for enzyme activity.

### Preparation of Working Solution

1. Add 50 µL of EK Yellow™ stock solution and 50 µL of Enterokinase Substrate stock solution into 5 mL of Assay Buffer; mix well to make Enterokinase (EK) working solution.

**Note:** The assay mixture is enough for one 96-well plate. It is not stable, use promptly.

### Experimental Protocol

BL	BL	TS	TS
EK1	EK1	...	...
EK2	EK2	...	...
EK3	EK3		
EK4	EK4		
EK5	EK5		
EK6	EK6		
EK7	EK7		

**Table 1.** Layout of enterokinase standards and test samples in a 96-well clear bottom microplate. EK = enterokinase standard (EK1 - EK7, 1.56 to 100 ng/mL); BL = blank control; TS = test sample.

Well	Volume	Reagent
EK1-EK7	50 µL	Serial Dilution (1.56 to 100 ng/mL)
BL	50 µL	Assay Buffer
TS	50 µL	Test Sample

**Table 2.** Reagent composition for each well of 96-well microplate.

1. Prepare enterokinase standards (EK), blank controls (BL), and test samples (TS) into a 96-well clear bottom microplate according to the layout provided in Table 1 and Table 2. For a 384-well plate, use 25  $\mu$ L of reagent per well instead of 50  $\mu$ L.
2. Add 50  $\mu$ L of EK working solution into each well of enterokinase standard, blank control, and test samples to make the total assay volume of 100  $\mu$ L/well. For a 384-well plate, add 25  $\mu$ L of EK working solution into each well instead, for a total volume of 50  $\mu$ L/well.
3. Incubate the reaction mixture at 37 °C for 30 - 60 minutes.
4. Monitor the absorbance increase with an absorbance plate reader with path check on at OD of 405 nm.

## Data Analysis

The reading (Absorbance) obtained from the blank standard well is used as a negative control. Subtract this value from the other standards' readings to obtain the base-line corrected values. Then, plot the standards' readings to obtain a standard curve and equation.

**For technical support contact information, visit:** [www.abcam.com/contactus](http://www.abcam.com/contactus)

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