

ab325364 – Human GLP1 (7-36) SimpleStep ELISA® Kit, Chemiluminescent

For the quantitative measurement of GLP1 (7-36) in human serum, plasma (citrate), plasma (heparin), and cell culture supernatant.

For research use only - not intended for diagnostic use.

Patent Pending

For overview, typical data and additional information please visit: www.abcam.com/ab325364

Storage and Stability: Store kit at 2-8°C immediately upon receipt. Refer to list of materials supplied for storage conditions of individual components. Observe the storage conditions for individual prepared components in the Standard Preparation and Reagent preparation sections.

Limitations: All data, except Typical Standard Curve and Sensitivity were collected using the colorimetric version of this kit (ab184857).

Materials Supplied

| Item | Quantity | Storage Condition |
|---|----------|-------------------|
| Human GLP1 (7-36) Capture Antibody 10X | 600 µL | +4°C |
| Human GLP1 (7-36) Detector Antibody 10X | 600 µL | +4°C |
| Human GLP1 (7-36) Lyophilized Recombinant Protein | 2 Vials | +4°C |
| Antibody Diluent CPI2 | 6 mL | +4°C |
| Sample Diluent NS | 50 mL | +4°C |
| Wash Buffer PT 10X | 20 mL | +4°C |
| ChemiHRP Reagent A | 3 mL | +4°C |
| ChemiHRP Reagent B | 3 mL | +4°C |
| SimpleStep Pre-Coated Black 96-Well Microplate | 96 Wells | +4°C |
| Plate Seal | 1 | +4°C |

Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully utilize this assay:

- Luminometer with the following settings: 0.5-1 second/well read time; summation mode (all wavelengths)
- Deionized water.
- Multi- and single-channel pipettes.
- Tubes for standard dilution.
- Plate shaker for all incubation steps.
- Optional: Phenylmethylsulfonyl Fluoride (PMSF) (or other protease inhibitors).

Reagent Preparation

Equilibrate all reagents to room temperature (18-25°C) prior to use. The kit contains enough reagents for 96 wells. The sample volumes below are sufficient for 48 wells (6 x 8-well strips); adjust volumes as needed for the number of strips in your experiment.

Prepare only as much reagent as is needed on the day of the experiment. Capture and Detector Antibodies have only been tested for stability in the provided 10X formulations.

1X Wash Buffer PT: Prepare 1X Wash Buffer PT by diluting Wash Buffer PT 10X with deionized water. To make 50 mL 1X Wash Buffer PT combine 5 mL Wash Buffer PT 10X with 45 mL deionized water. Mix thoroughly and gently.

Antibody Cocktail: Prepare Antibody Cocktail by diluting the capture and detector antibodies in Antibody Diluent CPI2. To make 3 mL of the Antibody Cocktail combine 300 µL 10X Capture Antibody and 300 µL 10X Detector Antibody with 2.4 mL Antibody Diluent CPI2. Mix thoroughly and **very** gently.

Lumi HRP Development Solution: Just prior to use, prepare Lumi HRP Development Solution by mixing equal volume of the ChemiHRP Reagent A and the ChemiHRP Reagent B. To make 3 mL of the Lumi HRP Development Solution combine 1.5 mL of ChemiHRP Reagent A and 1.5 mL of ChemiHRP Reagent B. Mix thoroughly and gently by inversion or slow pipetting (Avoid shaking or vortexing). Protect the prepared solution from light until use.

Standard Preparation

Always prepare a fresh set of standards for every use. Discard working standard dilutions after use as they do not store well. The following section describes the preparation of a standard curve for duplicate measurements (recommended).

1. Reconstitute the GLP1 (7-36) standard sample by adding the volume of Sample Diluent NS indicated on the protein vial label. Hold at room temperature for 10 minutes. Mix thoroughly and gently. This is the 1,000 pg/mL **Standard#1** Solution.
2. Label six tubes, Standards 1– 6.
3. Add 150 µL of Sample Diluent NS into numbers 2-6.
4. Use the **Stock Standard** to prepare the following dilution series. Standard #6 contains no protein and is the Blank control:

| Standard # | Dilution Sample | Volume to Dilute (µL) | Volume of Diluent (µL) | Starting Conc. (pg/mL) | Final Conc. (pg/mL) |
|------------|-------------------|-----------------------|------------------------|------------------------|---------------------|
| 1 | Standard#1 | 300 | 0 | 1,000 | 1,000 |
| 2 | Standard#1 | 75 | 150 | 1,000 | 333.3 |
| 3 | Standard#2 | 75 | 150 | 333.3 | 111.1 |
| 4 | Standard#3 | 75 | 150 | 111.1 | 37.0 |
| 5 | Standard#4 | 75 | 150 | 37.0 | 12.3 |
| 6 | Blank Control | 0 | 150 | 0 | 0 |

Sample Preparation

| Typical Sample Dynamic Range | |
|------------------------------|-------|
| Sample Type | Range |
| Serum* | ≤ 10% |
| Plasma – Citrate* | ≤ 10% |
| Plasma – Heparin* | ≤ 1% |
| Cell Culture Media* | ≤ 10% |

*Based on spiked sample

Serum Samples should be collected into a serum separator tube. After clot formation, centrifuge samples at 2,000 x g for 10 minutes and collect serum. Dilute samples at least 1:10 into Sample Diluent NS and assay. Store un-diluted serum at -20°C or below. Avoid repeated freeze-thaw cycles.

Plasma Collect plasma using citrate or heparin. Centrifuge samples at 2,000 x g for 10 minutes. Dilute citrate samples at least 1:10 and heparin samples at least 1:100 into Sample Diluent NS and assay. Store un-diluted plasma samples at -20°C or below for up to 3 months. Avoid repeated freeze-thaw cycles. Note: This kit is incompatible with plasma (EDTA) samples.

Cell Culture Supernatants Centrifuge cell culture media at 2,000 x g for 10 minutes to remove debris. Collect supernatants and assay. Or dilute samples at least 1:10 into Sample Diluent NS and assay. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

Plate Preparation

The 96 well plate strips included with this kit are supplied ready to use. It is not necessary to rinse the plate prior to adding reagents.

Unused plate strips should be immediately returned to the foil pouch containing the desiccant pack, resealed and stored at 4°C.

For each assay performed, a minimum of two wells must be used as the zero control.

For statistical reasons, we recommend each sample should be assayed with a minimum of two replicates (duplicates).

Differences in well absorbance or “edge effects” have not been observed with this assay.

Assay Procedure

Equilibrate all materials and prepared reagents to room temperature prior to use.

We recommend that you assay all standards, controls and samples in duplicate

1. Prepare all reagents, working standards, and samples as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, reseal and return to 4°C storage.
3. Add 50 µL of all sample or standard to appropriate wells.
4. Add 50 µL of the Antibody Cocktail to each well.
5. Seal the plate and incubate for 30 minutes at room temperature on a plate shaker set to 750 rpm.
6. Wash each well with 3 x 350 µL 1X Wash Buffer PT. Wash by aspirating or decanting from wells then dispensing 350 µL 1X Wash Buffer PT into each well. Wash Buffer PT should remain in wells for at least 30 seconds. Complete removal of liquid at each step is essential for good performance. After the last wash invert the plate and tap gently against clean paper towels to remove excess liquid.
7. Add 50 µL of prepared Lumi HRP Development Solution to each well and incubate for 1 minute in the dark on a plate shaker set to 750 rpm. Further optimization of incubation time vs signal strength can be performed if needed. Avoid introducing bubbles into the wells.
8. Measure the produced light of each well using a microplate luminometer with the following settings: 0.5-1 second/well read time in summation mode (all wavelengths). Relative light unit (RLU) readings may vary between luminometer models. It is recommended to configure instrument settings according to the manufacturer's specifications. Note: Relative light unit (RLU) values may change over the course of the 15-minute reading window.
9. Analyze the data as described below.

| | |
|-----------------------------|-----------------|
| Mode: | Luminescence |
| Instrument settings: | Endpoint |
| Detection Mode: | All wavelengths |
| Read Time: | 0.5-1 sec |
| Read: | Top |

Note For microplate readers with Pre-Read Optimization option, the Read Height as well as Microplate Optimization is recommended before the first read.

Download our ELISA guide for technical hints, results, calculation, and troubleshooting tips:

www.abcam.com/protocols/the-complete-elisa-guide

For technical support contact information, visit: www.abcam.com/contactus

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Additional information

ASSAY SPECIFICITY

This kit is designed for the quantification of human GLP1 (7-36).

The standard protein in this kit is human GLP1 (7-36), a fragment of GLP1 representing approximately 11% of the total length of GLP1.

Native signal was detected in serum sample types.

Spiked protein experiments were used to validate serum, plasma (citrate), plasma (heparin), and cell culture supernatant sample types.

Saliva, urine, milk, CSF, cell extract, and tissue extract samples have not been tested with this kit.

This kit is incompatible with plasma (EDTA) samples.

CROSS REACTIVITY

GLP-1 (9-36) recombinant protein was diluted at 10 ng/mL and assayed for cross reactivity. Signal was found at 7% from GLP-1 (7-36) protein assayed at the same concentration.

SPECIES REACTIVITY

Due to sequence similarity, this kit is predicted to cross react with multiples species such as mouse, rat, bovine, sheep, rabbit, pig and horse.

Other species reactivity not determined.

CALCULATION

- Preconfigured protocols are available when using SoftMax Pro software from Molecular Devices.
- Calculate the average chemiluminescence value for the blank control (zero) standards. Subtract the average blank control standard chemiluminescence value from all other chemiluminescence values.
- Create a standard curve by plotting the average blank control subtracted chemiluminescence value for each standard concentration (y-axis) against the target protein concentration (x-axis) of the standard. Use graphing software to draw the best smooth curve through these points to construct the standard curve.
- Note: Most chemiluminescence reader software or graphing software will plot these values and fit a curve to the data. A four-parameter curve fit (4PL) is often the best choice; however, other algorithms (e.g. linear, semi-log, log/log, 4-parameter logistic) can also be tested to determine if it provides a better curve fit to the standard values.
- Determine the concentration of the target protein in the sample by interpolating the blank control subtracted chemiluminescence values against the standard curve. Multiply the resulting value by the appropriate sample dilution factor, if used, to obtain the concentration of target protein in the sample.

- Samples generating chemiluminescence values greater than that of the highest standard should be further diluted and reanalyzed. Similarly, samples which measure at chemiluminescence values less than that of the lowest standard should be retested in a less dilute form.

TYPICAL DATA

Typical standard curve – data provided for demonstration purposes only. A new standard curve must be generated for each assay performed

| Standard Curve Measurements | | | |
|-----------------------------|-----------|-----------|-----------|
| Concentration (pg/mL) | RLU | | Mean RLU |
| | 1 | 2 | |
| 0 | 1,922 | 1,922 | 1,922 |
| 12.3 | 2,673 | 2,903 | 2,788 |
| 37.0 | 7,997 | 9,373 | 8,685 |
| 111.1 | 52,551 | 55,097 | 53,824 |
| 333.3 | 328,260 | 343,520 | 335,890 |
| 1,000 | 1,439,300 | 1,552,800 | 1,496,050 |

Table 1. Example of human GLP1 (7-36) standard curve in Sample Diluent NS. The GLP1 (7-36) standard curve was prepared as described in the Standard Preparation section. The table shows raw data values.

TYPICAL SAMPLE VALUES

Sensitivity:

The calculated minimal detectable dose (MDD) is 8.6 pg/mL. The MDD was determined by calculating the mean of zero standard replicates (n=16) and adding 2 standard deviations then extrapolating the corresponding concentration.

Recovery

Three concentrations of GLP1 (7-36) were spiked in duplicate to the indicated biological matrix to evaluate signal recovery in the working range of the assay.

| Sample Type | Average % Recovery | Range (%) |
|-------------------------|--------------------|-----------|
| 10% Serum | 109 | 105 - 112 |
| 10% Plasma (Citrate) | 115 | 107 - 123 |
| 1% Plasma (Heparin) | 110 | 104 - 112 |
| 10% Cell Culture Media* | 99 | 93 - 102 |

*Media is RPMI 1640 containing 10% fetal bovine serum.

Linearity of Dilution

Linearity of dilution is determined based on interpolated values from the standard curve. Linearity of dilution defines a sample concentration interval in which interpolated target concentrations are directly proportional to sample dilution.

Recombinant GLP1 (7-36) was spiked into the following biological samples in a 2-fold dilution series. Sample dilutions are made in Sample Diluent NS.

| Dilution Factor | Interpolated value | 10% Human Serum | 10% Human Plasma (Citrate) | 1% Human Plasma (Heparin) | 10% Cell Culture Media |
|-----------------|-------------------------|-----------------|----------------------------|---------------------------|------------------------|
| Undiluted | pg/mL | 258 | 261 | 265 | 238 |
| | % Expected value | 100 | 100 | 100 | 100 |
| 2 | pg/mL | 136 | 146 | 127 | 125 |
| | % Expected value | 106 | 112 | 96 | 105 |
| 4 | pg/mL | 69 | 73 | 68 | 61 |
| | % Expected value | 106 | 113 | 101 | 102 |
| 8 | pg/mL | 28 | 28 | 35 | 24 |
| | % Expected value | 87 | 84 | 105 | 80 |

Precision

Mean coefficient of variations of interpolated values of GLP1 (7-36) from three concentrations of human serum within the working range of the assay.

| | Intra-assay | Inter-assay |
|---------------|-------------|-------------|
| N= | 8 | 3 |
| CV (%) | 7.0 | 9.0 |

Download our ELISA guide for technical hints, results, calculation, and troubleshooting tips:

www.abcam.com/protocols/the-complete-elisa-guide

For technical support contact information, visit: www.abcam.com/contactus

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