

ab326733 – GFP SimpleStep ELISA® Kit – Extracellular, Chemiluminescent

For the quantitative measurement of GFP in cell culture supernatant.
For research use only - not intended for diagnostic use.
Patent pending.

For overview, typical data and additional information please visit: www.abcam.com/ab326733

Storage and Stability: Store kit at 2-8°C immediately upon receipt. Refer to list of materials supplied for storage conditions of individual components. Observe the storage conditions for individual prepared components in the Standard Preparation and Reagent preparation sections.

Limitations: All data, except typical standard curve and sensitivity were collected using the colorimetric version of this kit (ab316915).

Materials Supplied

Item	Quantity 1 x 96 tests	Storage Condition
GFP Capture Antibody 10X	600 µL	+4°C
GFP Detector Antibody 10X	600 µL	+4°C
GFP Lyophilized Recombinant Protein	2 Vials	+4°C
Antibody Diluent CPR2	6 mL	+4°C
Cell Extraction Enhancer Solution 50X	1 mL	+4°C
Sample Diluent NS	50 mL	+4°C
Wash Buffer PT 10X	20 mL	+4°C
ChemiHRP Reagent A	3 mL	+4°C
ChemiHRP Reagent B	3 mL	+4°C
SimpleStep Pre-Coated Black 96-Well Microplate	96 Wells	+4°C
Plate Seal	1	+4°C

Materials Required, Not Supplied

These materials are not included in the kit, but will be required to successfully utilize this assay:

Luminometer capable of reading strip plates with the following settings: 0.5-1 second/well read time; summation mode (all wavelengths).

Deionized water.

Multi- and single-channel pipettes.

Tubes for standard dilution.

Orbital microplate shaker for all incubation steps: capable of 750 rpm shaking speed.

Optional: Phenylmethylsulfonyl Fluoride (PMSF) (or other protease inhibitors).

Reagent Preparation

Equilibrate all reagents to room temperature (18-25°C) prior to use. The kit contains enough reagents for 96 wells. The sample volumes below are sufficient for 48 wells (6 x 8-well strips); adjust volumes as needed for the number of strips in your experiment.

Prepare only as much reagent as is needed on the day of the experiment. Capture and Detector Antibodies have only been tested for stability in the provided 10X formulations.

The provided Cell Extraction Enhancer Solution 50X may precipitate when stored at + 4°C. To dissolve, warm briefly at + 37°C and mix gently. The Cell Extraction Enhancer Solution 50X can be stored at room temperature to avoid precipitation.

1X Wash Buffer PT: Prepare 1X Wash Buffer PT by diluting Wash Buffer PT 10X with deionized water. To make 50 mL 1X Wash Buffer PT combine 5 mL Wash Buffer PT 10X with 45 mL deionized water. Mix thoroughly and gently.

Sample Diluent NS + 1X Enhancer: Prepare Sample Diluent NS + 1X Enhancer by combining Sample Diluent NS and 50X Cell Extraction Enhancer Solution. To make 5 mL Sample Diluent NS + 1X Enhancer, combine 4.9 mL Sample Diluent NS and 100 µL Cell Extraction Enhancer Solution 50X. Mix thoroughly and gently.

Antibody Cocktail: Prepare Antibody Cocktail by diluting the capture and detector antibodies in Antibody Diluent CPR2. To make 3 mL of the Antibody Cocktail combine 300 µL 10X Capture Antibody and 300 µL 10X Detector Antibody with 2.4 mL Antibody Diluent CPR2. Mix thoroughly and gently.

Lumi HRP Development Solution: Just prior to use, prepare Lumi HRP Development Solution by mixing equal volume of the ChemiHRP Reagent A and the ChemiHRP Reagent B. To make 3 mL of the Lumi HRP Development Solution combine 1.5 mL of ChemiHRP Reagent A and 1.5 mL of ChemiHRP Reagent B. Mix thoroughly and gently by inversion or slow pipetting (Avoid shaking or vortexing). Protect the prepared solution from light until use.

Standard Preparation

Always prepare a fresh set of standards for every use. Discard working standard dilutions after use as they do not store well. The following section describes the preparation of a standard curve for duplicate measurements (recommended).

1. Reconstitute the GFP standard sample by adding the volume of **Sample Diluent NS + 1X Enhancer** indicated on the protein vial label. Hold at room temperature for 10 minutes. Mix thoroughly and gently. This is the 9.6 ng/mL **Stock Standard** Solution.
2. Label nine tubes, Standards 1– 9.
3. Add 247 µL of Sample Diluent NS + 1X Enhancer into tube number 1 and 150 µL of Sample Diluent NS + 1X Enhancer into numbers 2-9.
4. Use the **Stock Standard** to prepare the following dilution series. Standard #9 contains no protein and is the Blank control:

Standard #	Dilution Sample	Volume to Dilute (µL)	Volume of Diluent (µL)	Starting Conc. (pg/mL)	Final Conc. (pg/mL)
1	Stock Standard	57	247	9,600	1,800
2	Standard#1	75	150	1,800	600
3	Standard#2	75	150	600	200
4	Standard#3	75	150	200	66.7
5	Standard#4	75	150	66.7	22.2
6	Standard#5	75	150	22.2	7.4
7	Standard#6	75	150	7.4	2.5
8	Standard#7	75	150	2.5	0.8
9	Blank Control	0	150	0	0

Sample Preparation

Typical Sample Dynamic Range	
Sample Type	Range
HeLa Cell Supernatant	≤ 50%
Neuro 2A Cell Supernatant	≤ 50%
Mouse Lung Tissue Supernatant	≤50%

Note: This kit is compatible with samples containing 0.1% SDS. If sample contains higher concentration of SDS, dilute initially the sample with Sample Diluent NS without Enhancer to lower SDS concentration to 0.1%. Following dilutions can be done in **Sample Diluent NS + 1X Enhancer**.

Cell Culture Supernatants Centrifuge cell culture media at 2,000 x g for 10 minutes to remove debris. Collect supernatants. Dilute samples at least 1:2 into **Sample Diluent NS + 1X Enhancer** and assay. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

Plate Preparation

The 96 well plate strips included with this kit are supplied ready to use. It is not necessary to rinse the plate prior to adding reagents. Unused plate strips should be immediately returned to the foil pouch containing the desiccant pack, resealed and stored at 4°C. For each assay performed, a minimum of two wells must be used as the zero control. For statistical reasons, we recommend each sample should be assayed with a minimum of two replicates (duplicates). Differences in well absorbance or “edge effects” have not been observed with this assay.

Assay Procedure

Equilibrate all materials and prepared reagents to room temperature prior to use.
We recommend that you assay all standards, controls and samples in duplicate.

1. Prepare all reagents, working standards, and samples as directed in the previous sections.
2. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, reseal and return to 4°C storage.
3. Add 50 µL of all sample or standard to appropriate wells.
4. Add 50 µL of the Antibody Cocktail to each well.
5. Seal the plate and incubate for 30 minutes at room temperature on a plate shaker set to 750 rpm.
6. Wash each well with 3 x 350 µL 1X Wash Buffer PT. Wash by aspirating or decanting from wells then dispensing 350 µL 1X Wash Buffer PT into each well. Wash Buffer PT should remain in wells for at least 30 seconds. Complete removal of liquid at each step is essential for good performance. After the last wash invert the plate and tap gently against clean paper towels to remove excess liquid.
7. Add 50 µL of prepared Lumi HRP Development Solution to each well and incubate for 1 minute in the dark on a plate shaker set to 750 rpm. Further optimization of incubation time vs signal strength can be performed if needed. Avoid introducing bubbles into the wells.
8. Measure the produced light of each well using a microplate luminometer with the following settings: 0.5-1 second/well read time in summation mode (all wavelengths). Relative light unit (RLU) readings may vary between luminometer models. It is recommended to configure instrument settings according to the manufacturer's specifications. Note: Relative light unit (RLU) values may change over the course of the 15-minute reading window.
9. Analyze the data as described below.

Mode:	Luminescence
Instrument settings:	Endpoint
Detection Mode:	All wavelengths
Read Time:	0.5-1 sec
Read:	Top

Note For microplate readers with Pre-Read Optimization option, the Read Height as well as Microplate Optimization is recommended before the first read.

Download our ELISA guide for technical hints, results, calculation, and troubleshooting tips:
<https://www.abcam.com/en-us/technical-resources/guides/elisa-guide>

Technical Support

Copyright © 2026 Abcam, All Rights Reserved. The Abcam logo is a registered trademark. All information / detail is correct at time of going to print.
Version 1a | 2026-04-13

For all technical or commercial enquiries please go to:
<https://www.abcam.com/en-us/contact-us>
<https://www.abcam.cn/contact-us> (China)
<https://www.abcam.co.jp/contact-us> (Japan)

ab326733 – GFP SimpleStep ELISA® Kit – Extracellular, Chemiluminescent

Additional information

ASSAY SPECIFICITY

This kit is designed for the quantification of GFP.

The standard protein in this kit is full length enhanced GFP (eGFP).

This kit recognizes the GFP, eGFP, as well as the SDS-denatured eGFP.

500 pg/mL of recombinant GFP was tested for reactivity. 92% reactivity was observed.

600 pg/mL of recombinant SDS-denatured EGFP was tested for reactivity. 89% reactivity was observed.

Spiked protein experiments were used to validate human and mouse cell and tissue culture supernatant sample types.

50% HeLa, Neuro 2A, and mouse lung cell culture supernatants were measured in duplicate. All values were below the detectable range of the assay.

Serum, plasma (citrate), plasma (EDTA), plasma (heparin), urine, milk, saliva, CSF, cell extract, and tissue extract samples have not been tested with this kit.

For the measurement of GFP in cell and tissue extract sample types, use GFP Chemiluminescent ELISA kit ab326732.

CROSS REACTIVITY

6,000 pg/mL of recombinant YFP and 6,000 pg/mL of recombinant mCherry were tested for cross reactivity. No cross reactivity was observed.

INTERFERENCE

6,000 pg/mL of recombinant YFP and 6,000 pg/mL of recombinant mCherry were tested for interference with 600 pg/mL of recombinant GFP. No interference was observed.

CALCULATION

- Preconfigured protocols are available when using SoftMax Pro software from Molecular Devices.
- Calculate the average chemiluminescence value for the blank control (zero) standards. Subtract the average blank control standard chemiluminescence value from all other chemiluminescence values.
- Create a standard curve by plotting the average blank control subtracted chemiluminescence value for each standard concentration (y-axis) against the target protein concentration (x-axis) of the standard. Use graphing software to draw the best smooth curve through these points to construct the standard curve.
- Note: Most chemiluminescence reader software or graphing software will plot these values and fit a curve to the data. A four-parameter curve fit (4PL) is often the best choice; however, other algorithms (e.g. linear, semi-log, log/log, 4-parameter logistic) can also be tested to determine if it provides a better curve fit to the standard values.
- Determine the concentration of the target protein in the sample by interpolating the blank control subtracted chemiluminescence values against the standard curve. Multiply the resulting value by the appropriate sample dilution factor, if used, to obtain the concentration of target protein in the sample.
- Samples generating chemiluminescence values greater than that of the highest standard should be further diluted and reanalyzed. Similarly, samples which measure at chemiluminescence values less than that of the lowest standard should be retested in a less dilute form.

TYPICAL DATA

Typical standard curve – data provided for demonstration purposes only. A new standard curve must be generated for each assay performed.

Standard Curve Measurements			
Concentration (pg/mL)	RLU		Mean RLU
	1	2	
0	419	591	505
0.82	1,358	1,450	1,404
2.47	3,261	3,172	3,217
7.41	11,299	10,897	11,098
22.2	39,921	36,248	38,085
66.7	131,660	123,440	127,550
200	372,980	443,410	408,195
600	1,115,400	1,163,800	1,139,600
1,800	4,558,000	4,023,100	4,290,550

Table 1. Example of GFP standard curve in **Sample Diluent NS + 1X Enhancer**. The GFP standard curve was prepared as described in the Standard Preparation section. The table shows raw data values.

TYPICAL SAMPLE VALUES

Sensitivity:

The calculated minimal detectable dose (MDD) is 0.29 pg/mL. The MDD was determined by calculating the mean of zero standard replicates (n=16) and adding 2 standard deviations then extrapolating the corresponding concentration.

Recovery

Three concentrations of eGFP were spiked in duplicate to the indicated biological matrix to evaluate signal recovery in the working range of the assay.

Sample Type	Average % Recovery	Range (%)
50% HeLa Cell Culture Supernatant*	98	94 - 104
50% Neuro 2A Cell Culture Supernatant*	101	91 - 108
50% Mouse Lung Tissue Culture Supernatant**	86	79 - 93

*Media is DMEM containing 10% fetal bovine serum

** Media is RPMI 1640 containing 10% fetal bovine serum

Linearity of Dilution

Linearity of dilution is determined based on interpolated values from the standard curve. Linearity of dilution defines a sample concentration interval in which interpolated target concentrations are directly proportional to sample dilution.

Recombinant eGFP was spiked into the following biological samples in a 2-fold dilution series. Sample dilutions are made in **Sample Diluent NS + 1X Enhancer**.

Dilution Factor	Interpolated value	50% HeLa Cell Supernatant	50% Neuro 2A Cell Supernatant	50% Mouse Lung Tissue Supernatant
Undiluted	pg/mL	237.0	279.8	287.0
	% Expected value	100%	100%	100%
2	pg/mL	111.2	164.3	146.1
	% Expected value	94%	117%	102%
4	pg/mL	57.6	82.7	69.9
	% Expected value	97%	118%	97%
8	pg/mL	33.0	39.6	37.7
	% Expected value	111%	113%	105%
16	pg/mL	16.8	16.6	15.5
	% Expected value	113%	95%	86%

Precision

Mean coefficient of variations of interpolated values of GFP from two concentrations of HeLa cell culture supernatant within the working range of the assay.

	Intra-assay	Inter-assay
N=	8	3
CV (%)	8.8	7.4

Download our ELISA guide for technical hints, results, calculation, and troubleshooting tips:

<https://www.abcam.com/en-us/technical-resources/guides/elisa-guide>

Technical Support

Copyright © 2026 Abcam, All Rights Reserved. The Abcam logo is a registered trademark. All information / detail is correct at time of going to print.

Version 1a | 2026-04-13

For all technical or commercial enquiries please go to:

<https://www.abcam.com/en-us/contact-us>

<https://www.abcam.cn/contact-us> (China)

<https://www.abcam.co.jp/contact-us> (Japan)