

ab327364 – Ultra Lipofuscin Autofluorescence Blocker

This product effectively suppresses lipofuscin and other tissue autofluorescence with superior water solubility and enhanced quenching capability.

For research use only - not intended for diagnostic use.

For overview, typical data and additional information please visit: www.abcam.com/ab327364

Storage and Stability: Store product at -20°C in the dark immediately upon receipt.

Materials Supplied

Item	Quantity	Storage Condition
Ultra Lipofuscin Autofluorescence Blocker	1 mL	-20°C

Protocol Summary

1. Perform fixation, deparaffinization, and antigen retrieval of tissue sections as per your standard protocol.
2. Add Ultra Lipofuscin Autofluorescence Blocker working solution to the sample.
3. Incubate the sample at room temperature for 5 to 10 minutes.
4. Apply the mounting medium and examine the sample using fluorescence microscopy.

ΔNote:

Thaw the Ultra Lipofuscin Autofluorescence Blocker at room temperature. If any precipitates are observed, gently heat the vial at 37°C for 5 to 10 minutes before preparing the working solution.

Preparation of Working Solution

To prepare the Ultra Lipofuscin Autofluorescence Blocker working solution (1X), dilute 25 µL of the 40X DMSO stock solution in 1 mL of PBS. Mix thoroughly.

ΔNotes:

- Protect the working solution from light by covering it with foil or storing it in a dark location.
- For optimal results, use this solution within a few hours of preparation.
- Prepare 100 to 200 µL of Ultra Lipofuscin Autofluorescence Blocker working solution (1X) for each tissue section that will be treated.

Sample Experimental Protocol

Pretreatment with Ultra Lipofuscin Autofluorescence Blocker

1. Perform fixation, deparaffinization, and/or antigen retrieval of the tissue sections according to your standard protocols.
2. Wash slides with PBS.
3. Remove the PBS, then place the tissues in a humidified slide chamber.

4. Add 100 to 200 µL of Ultra Lipofuscin Autofluorescence Blocker working solution (1X) to each tissue sample, ensuring the solution fully covers the samples.
5. Incubate the slides at room temperature for 5 to 10 minutes.
ΔNote: The incubation time may vary based on your specific application and can be adjusted for optimal results.
6. Wash the slides two times with PBS.
7. Perform immunofluorescence staining with antibodies according to your recommended protocol.

ΔNote: Avoid using buffers containing detergents during blocking, antibody incubation, or washing steps. If detergents are necessary, use them according to the post-treatment protocol.

8. Cover the slides using any aqueous-based fluorescence anti-fade mounting medium.
ΔNote: Avoid using organic-based mounting mediums, as they are incompatible with Ultra Lipofuscin Autofluorescence Blocker.

Posttreatment with Ultra Lipofuscin Autofluorescence Blocker

1. Perform fixation, deparaffinization, and/or antigen retrieval of the tissue sections according to your standard protocols.
2. Perform immunofluorescence staining with antibodies according to your recommended protocol.

ΔNote: Avoid using buffers containing detergents during blocking, antibody incubation, or washing steps. If detergents are necessary, use them according to the post-treatment protocol.

3. Wash the slides with PBS.
4. Remove the PBS, then place the tissues in a humidified slide chamber.
5. Add 100 to 200 µL of Ultra Lipofuscin Autofluorescence Blocker working solution (1X) to each tissue sample, ensuring the solution fully covers the samples.
6. Incubate the slides at room temperature for 5 to 10 minutes.
ΔNote: The incubation time may vary based on your specific application and can be adjusted for optimal results.

7. Wash the slides two times with PBS.
8. Cover the slides using any aqueous-based fluorescence anti-fade mounting medium.
ΔNote: Avoid using organic-based mounting mediums, as they are incompatible with Ultra Lipofuscin Autofluorescence Blocker.

Technical Support

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