Immunoprecipitation Protocol using GFP-Selector Affinity Resins.

Immunoprecipitation of fusion proteins from cell extracts using ab328083 GFP-Selector Affinity Resin (4% cross-linked agarose) and ab328084 GFP-Selector Affinity Resin (4% cross-linked magnetic agarose).

Storage: Store product at 2-8°C immediately upon receipt. **DO NOT FREEZE.**

Materials Required, Not Supplied

These materials are not included, but will be required to successfully utilize these products:

Lysis buffer
Washing buffer
Tris-buffered saline (TBS) pH 7.4
2x SDS sample buffer

∆***Note:*** *Selector resins are compatible with most common Lysis/Washing buffers (e.g. RIPA).*

Protocol using Mini Spin Columns

We recommend using Mini Spin Columns for immunoprecipitation experiments. An alternative batch protocol is provided in the next section.

1. Prepare native cell lysates (0.2 to 1.5 ml volume) according to established protocols.
For mammalian cells, we recommend using 10^6-10^8 cells per experiment.
2. Clarify lysate by centrifugation C for 10 min at > 14000 x g and 4 °C.
Take sample for further analysis (Input fraction)
3. Equilibrate GFP Selector Resin
 a. Resuspend GFP Selector Resin.
 b. Transfer 20 µl slurry (10 µl packed beads) into a clean 1.5 ml reaction tube.
 c. Add 1 ml lysis buffer.
 d. Centrifuge for 1 min at 1000 x g and carefully remove supernatant.
 e. Repeat steps c-d once.
4. Add clarified lysate from step 2 to equilibrated GFP Selector Resin obtained in step 3.
5. Incubate 1 h at 4 °C with head-over-tail rotation.
6. Sediment beads by centrifugation for 1 min at 1000 x g and 4 °C.
Take sample from supernatant for further analysis (Non-bound fraction)
7. Washing
 a. Carefully remove supernatant.
 b. Resuspend beads in 1 ml Lysis buffer
 c. Centrifuge for 1 min at 1000 x g.
 d. Remove Supernatant
8. Transfer
 a. Remove bottom plug from Mini Spin Column. Place column in 2 ml reaction tube.
 b. Resuspend beads in 200 µl Lysis buffer, transfer suspension to Mini Spin column
 c. Wash out beads sticking to tube with 200 µl Lysis buffer and transfer to column.
 d. Centrifuge column for 1 min at 1000 x g, discard flow-through.
9. Wash twice with 400 µl Washing buffer, centrifuge for 1 min at 1000 x g.
10. Wash once with 400 µl TBS, centrifuge for 1 min at 3000 x g.
11. Attach bottom plug and place Mini Spin Column in a clean 1.5 ml reaction tube
12. Resuspend GFP Selector resin in 50 µl 2x SDS sample buffer
13. Heat Mini Spin Column to 95 °C for 2 min.
14. Remove bottom plug and centrifuge for 1 min at 3000 x g.
Boil collected eluate for 5 min at 95 °C and analyze by SDS-PAGE.

Batch Protocol

Due to the more effective washing steps, we recommend using Protocol using Mini Spin Columns for immunoprecipitation experiments (see previous section).

1. Prepare native cell lysates (0.2 to 1.5 ml volume) according to established protocols.
For mammalian cells, we recommend using 10^6-10^8 cells per experiment.
2. Clarify lysate by centrifugation for 10 min at > 14000 x g and 4 °C.
Take sample for further analysis (Input fraction)
3. Equilibrate GFP Selector Resin
 a. Resuspend GFP Selector Resin.
 b. Transfer 20 µl slurry (10 µl packed beads) into a clean 1.5 ml reaction tube.
 c. Add 1 ml lysis buffer.
 d. Centrifuge for 1 min at 1000 x g and carefully remove supernatant.
 e. Repeat steps c-d once.
4. Add clarified lysate from step 2 to equilibrated GFP Selector Resin obtained in step 3.
5. Incubate 1 h at 4 °C with head-over-tail rotation.
6. Sediment beads by centrifugation for 1 min at 1000 x g and 4 °C.
Take sample from supernatant for further analysis (Non-bound fraction)
7. Washing
 a. Carefully remove supernatant.
 b. Resuspend beads in 1 ml Lysis buffer
 c. Centrifuge for 1 min at 1000 x g.
 d. Wash beads 2-3 times with Washing buffer
 e. Wash beads once with TBS
8. Transfer beads in clean 1.5 ml reaction tube.
9. Centrifuge for 1 min at 3000 x g.
10. Carefully and completely remove supernatant
11. Resuspend GFP Selector resin in 50 µl 2x SDS sample buffer
12. Heat for 5 min to 95 °C.
13. Centrifuge for 1 min at 3000 x g.
Collect supernatant and analyze by SDS-PAGE.

Technical Support

Copyright © 2026 Abcam, All Rights Reserved. The Abcam logo is a registered trademark. All information / detail is correct at time of going to print.
Version 1 | 2026-09-28

For all technical or commercial enquiries please go to:
https://www.abcam.com/en-us/contact-us
https://www.abcam.cn/contact-us (China)
https://www.abcam.co.jp/contact-us (Japan)